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(Νόμος 5343/32, άρθρο 202 παρ. 2)

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3. M. Dardioti, E. Dardiotis, K. Aggelakis, K. Paterakis, A. Komnos, A. Karantanas, A.Tasiou, G. Xiromerissiou, G. Hadjigeorgiou: **Interleukin 2 (+114) gene polymorphism and traumatic brain injury**. Sixteenth Meeting of the European Neurological Society, 27-31 May 2006, Lausanne, Switzerland. Journal of Neurology, Volume 253, Suppl.2, May 2006: II/53.

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Abstract	. 12
	. 15
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μ	. 34
	. 55
-1 (IL-1)	. 70
-2 (IL-2)	. 84
-6 (IL-6)	. 92
	. 100
μ	. 101
μ	. 102
	. 108
μ	. 140
	. 166
	. 180

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 μ μ μ μ
 μ μ μ μ μ -
 μ μ IL-1 (-511C/T, rs16944), IL-1 (+3953C/T, rs1143634), IL-1 (-
 889C/T, rs1800587), IL-1RA (VNTR 2 , rs2234663), IL-
 2(+114G/T, rs2069763), IL-2(-384 /G, rs2069762) IL-6(-174G/C, rs1800795).
 : μ 365 μ (60
 [16.4%]). μ μ
 μ μ , μ μ μ
 . μ μ
 μ .
 μ DNA μ μ ,
 μ μ μ PCR/RFLP μ .
 Hardy-Weinberg μ Fisher exact test.
 μ μ μ
 ² . μ
 μ μ , μ μ
 . μ μ μ
 μ μ μ μ
 μ μ μ μ
 Bonferroni: μ μ μ
 p=0.007 (0.05/7=0.007)
 μ : μ μ
 μ μ IL-1 (-511C/T), IL-1 (+3953C/T), IL-1 (-
 889C/T), IL-2 (-384T/G) IL-6(-174G/C).
 μ μ IL-2(+114G/T)
 μ μ . μ ,

μ μ μ μ GCS 3-8 μ μ
 GCS 13-15 μ μ (μ [μ
 μ] 0.57 [0.38-0.80], $p=0.002$) μ
 μ μ (0.53 [0.31-0.81], $p=0.009$)
 μ μ μ (2.32 [1.18-5.12], $p=0.02$).
 μ μ IL-1RA (VNTR)
 μ 2 4.5 μ μ -
 μ μ (4.57 [1.67-12.96], $p=0.004$). μ μ -
 μ (0.26 [0.14-
 0.71], $p=0.003$). , μ 2 μ
 : GCS 3-8 μ
 GCS 9-12 (μ 0.32 [0.12-0.90], $p=0.030$ μ
 0.32 [0.16-0.83], $p=0.02$) GCS 3-8 μ
 GCS 13-15 (μ 0.37 [0.19-0.78], $p=0.02$ μ
 0.45 [0.25-0.91], $p=0.017$). , 6μ ,
 μ 2 μ :
 μ 0.39 [0.19-0.89], $p=0.011$ μ μ 0.43 [0.19-
 0.86], $p=0.035$.
 μ μ
 IL-1 (IL-1 -889C/T, IL-1 +3953C/T, IL-1 -511C/T, IL-1RA VNTR) IV
 (1-1-2-2) μ μ
 (0.16 (0.05-0.51), $p=0.0005$) μ μ (0.40 (0.20-0.78),
 $p=0.0058$). , VIII (2-2-2-1) μ μ
 (9.79 (2.74-34.98), $p=2.64*10^{-5}$),
 VI (2-1-1-1) μ μ μ 6μ
 3.01 [1.59-5.69], $p=0.0004$).
 μ μ IL-2 (IL-2 +114G/T IL-2 -384T/G)
 II (2-1) μ μ (0.55
 (0.39-0.78), $p=0.00087$).
 μ μ : μ μ
 μ μ ,
 μ μ μ . ,
 μ
 μ μ μ .

ABSTRACT

Introduction: Traumatic Brain Injury (TBI) triggers inflammatory processes the extent of which determines the degree of acute and chronic neuronal cell death and patient's clinical outcome. Interleukins, which are the main mediators of inflammation, were found to play a central role in the pathophysiology of TBI. We conducted a prospective study to investigate possible influence of genetic polymorphisms of the interleukin genes in the clinical severity, 6 months outcome and hemorrhagic events in patients with TBI. In particular, we tested the IL-1 (-511C/T, rs16944), IL-1 (+3953C/T, rs1143634), IL-1 (-889C/T, rs1800587), IL-1RA (VNTR, rs2234663), IL-2(+114G/T, rs2069763), IL-2(-384T/G), rs2069762) and IL-6(-174G/C, rs1800795) polymorphisms.

Patient-Methods: 365 (women=60 [16.4%]) prospectively recruited patients with TBI were evaluated. Initial neurological status and six-month outcome were assessed by means of the Glasgow Coma Score and Glasgow Outcome Scale respectively. Based on computed tomography findings at admission patients were stratified according to the presence and type of haemorrhagic event.

DNA was extracted using the salting out method, while SNPs genotypes were determined using standard PCR/RFLP methods

The Hardy–Weinberg equilibrium was assessed by means of the Fisher's exact test. For comparisons between categorical variables we used the χ^2 test. Multivariable logistic regression analysis was used to control for possible confounding factors. Analysis for dominant recessive and allele contrast models of inheritance was performed. Linkage disequilibrium between SNPs was examined and haplotype estimation was performed by an expectation-maximization algorithm. Control for multiple comparisons was performed using Bonferroni's correction for 7 polymorphisms. The level of significance (p) was equal to 0.007 ($0.05/7=0.007$).

Results: No statistically significant difference was found for IL-1 (-511C/T), IL-1 (+3953C/T), IL-1 (-889C/T), IL-2 (-384T/G) and IL-6(-174G/C) polymorphisms.

Allele T of IL-2(+114G/T) polymorphism was associated with more severe clinical presentation at admission. There was a significant association between patients with GCS 3-8 and those with GCS 13-15 when considering allele contrast model (odds ratios [confidence interval] 0.57 [0.38-0.80], $p=0.002$) and a trend for association for dominant model (0.53 [0.31-0.81], $p=0.009$) and recessive model of inheritance (2.32 [1.18-5.12], $p=0.02$).

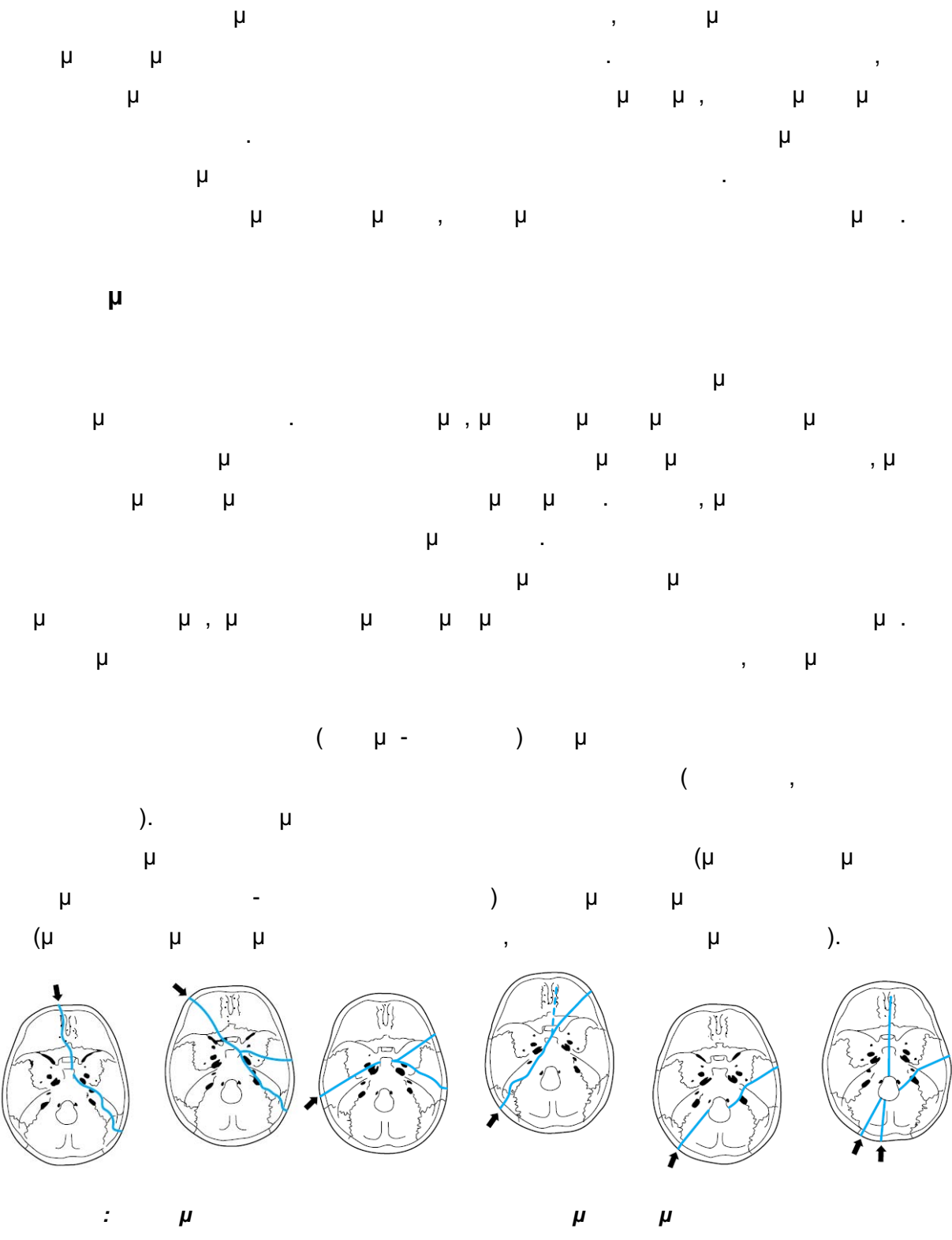
Compared to noncarriers, IL-1RA allele 2 carriers had 4.5 times higher odds of having cerebral hemorrhages after TBI (4.57 [1.67-12.96], $p=0.004$). Similar results

were obtained when comparing allele contrast frequencies (0.26 [0.14-0.71], $p=0.003$). In allele 2 carriers there was also a trend towards severe clinical presentation at admission: GCS at admission 3-8 vs GCS 9-12 (dominant genetic model 0.32 [0.12-0.90], $p=0.030$ and allele contrast model 0.32 [0.16-0.83], $p=0.02$) and GCS at admission 3-8 vs GCS 13-15 (dominant model 0.37 [0.19-0.78], $p=0.02$ and allele contrast model 0.45 [0.25-0.91], $p=0.017$). Finally, an opposite trend towards favourable 6 month's outcome was found for IL-1RA allele 2 (dominant model 0.39 [0.19-0.89], $p=0.011$ and allele contrast genetic model 0.43 [0.19-0.86], $p=0.035$).

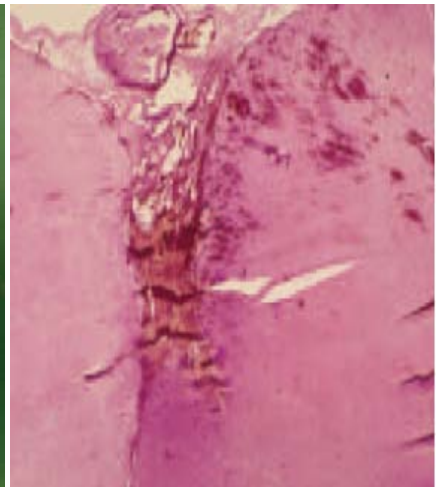
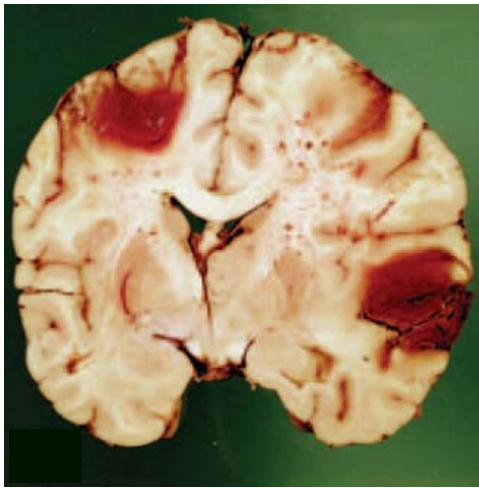
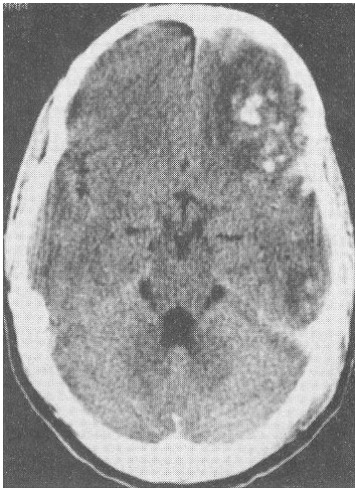
Haplotype analysis of IL-1 SNPs (IL-1 -889C/T, IL-1 +3953C/T, IL-1 -511C/T, IL-1RA VNTR) revealed that haplotype IV (1-1-2-2) was associated with increased risk for more severe clinical presentation at admission [0.16 (0.05-0.51), $p=0.0005$] and hemorrhagic events [0.40 (0.20-0.78), $p=0.0058$]. Haplotype VIII (2-2-2-1) was associated with higher likelihood for milder clinical presentation at admission [9.79 (2.74-34.98), $p=2.64 \times 10^{-5}$], while haplotype VI (2-1-1-1) was linked to unfavourable 6month outcome [3.01 (1.59-5.69), $p=0.0004$]. Haplotype analysis of IL-2 SNPs (IL-2 +114G/T, IL-2 -384T/G) revealed that haplotype II (2-1) was associated with increased risk for more severe clinical presentation at admission [0.55 (0.39-0.78), $p=0.00087$].

Conclusions: Our study provides evidence of an implication of interleukins genetic polymorphisms in the clinical presentation at admission, 6 months outcome and hemorrhagic events of TBI patients. However, further studies are needed to confirm these associations and investigate underlying pathogenetic mechanisms.

μ .) . μ μ μ (μ
 μ . _____ (concussion) μ
 , μ μ μ
 _____ (contusion)
 μ μ μ .
 μ coup
 , μ μ μ contracoup
 [2]. μ μ
 μ , μ μ
 μ , (shearing injury), μ .
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 μ μ , μ μ μ , μ μ μ .



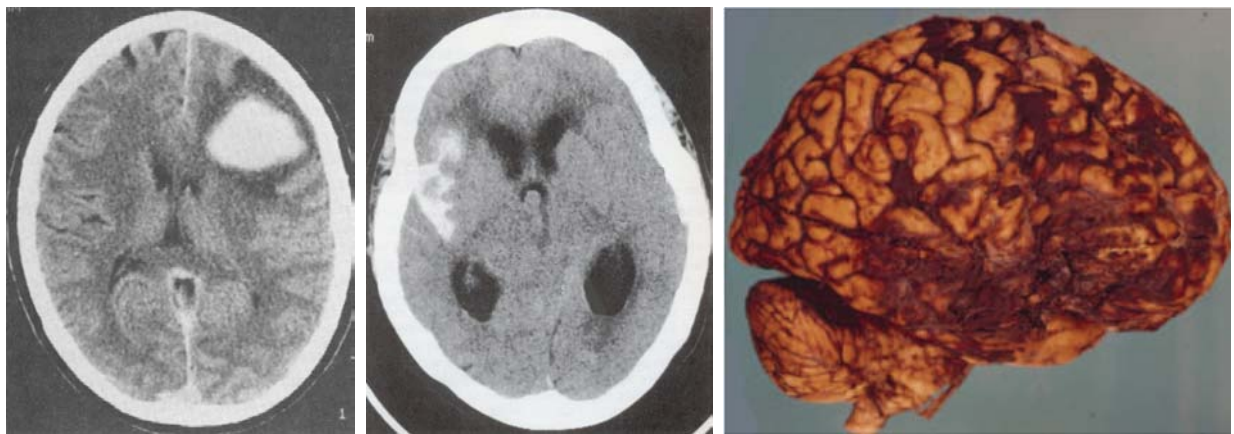
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μ μ μ μ μ μ μ μ [4].
 μ μ , μ μ μ μ μ μ μ μ « »
 μ μ μ μ μ μ μ μ .



μ (,) μ () μ μ -

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 (μ μ , μ
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 (μ μ μ),
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 μ μ μ μ μ μ μ μ

<90mm Hg)

(μ Battle),

(GCS: Glasgow Coma Scale)

3

(1) (2) (3)

(GCS<8:

μ μ (GCS)	
μ μ	μ 4
	μ 3
	μ 2
	μ 1
	μ 5
	μ 4
	μ 3
	μ 2
	μ 1
	μ 6
	μ 5
	μ () 4
	μ 3
	μ 2
	μ 1

: μ μ μ μ 3(μ) μ 15.

μ
 μ : μ μ μ
 μ . μ μ $\mu\mu$ μ μ .
 μ μ μ μ μ μ μ μ
 μ , μ , μ , $\mu\mu$
 μ μ μ μ μ μ μ μ μ

μ	(GOS)
(G):	-
$\mu\mu$.	μ
(MD): (μ) .	μ .
μ , μ , μ , $\mu\mu$ μ μ	μ μ μ μ
(SD): (μ) .	μ .
(V):	
(D)	

μ : μ μ μ
 μ 6μ μ μ .

, μ μ μ -
 , μ , (μ) ,
 μ (μ -
) () . μ
 Becker . [7], -
 , μ : : 32%, -
 / : 11%, μ / : 57%. -
 μ .
 μ μ μ .
 μ μ μ
 μ .
 μ μ
 , μ ,
 μ (punch-drunk boxer's syndrome), μ -
 μ μ . μ -
 μ μ μ
 . μ μ
 80% μ μ 3 μ 4,
 μ 54% GCS 5 μ 7, 27% μ μ (11-
 8 μ 10 6% μ μ 15).
 μ μ μ
 μ μ 65%
 μ CT (mass lesion) 82% μ μ -
 . μ μ μ 50% μ .
 60%. μ μ μ
 μ , (. .) .
 , μ .
 μ , μ .

[16-18].
 (Ca²⁺).
 ATP (adenosine triphosphate)
 Ca²⁺. NMDA
 (N-methyl-D-aspartate) Ca²⁺
 [19-22].
 Ca²⁺
 Ca²⁺
 [23].
 [24-26].
 N-methyl-d-aspartate
 metabotropic Kainate amino-3-hydroxy-5-
 methy-4-isoxazole propionate

μ
 . μ μ μ μ
 , μ [20-22]. ,
 μ μ μ μ μ -
 . μ (50
 μ
), μ
 μ [27].

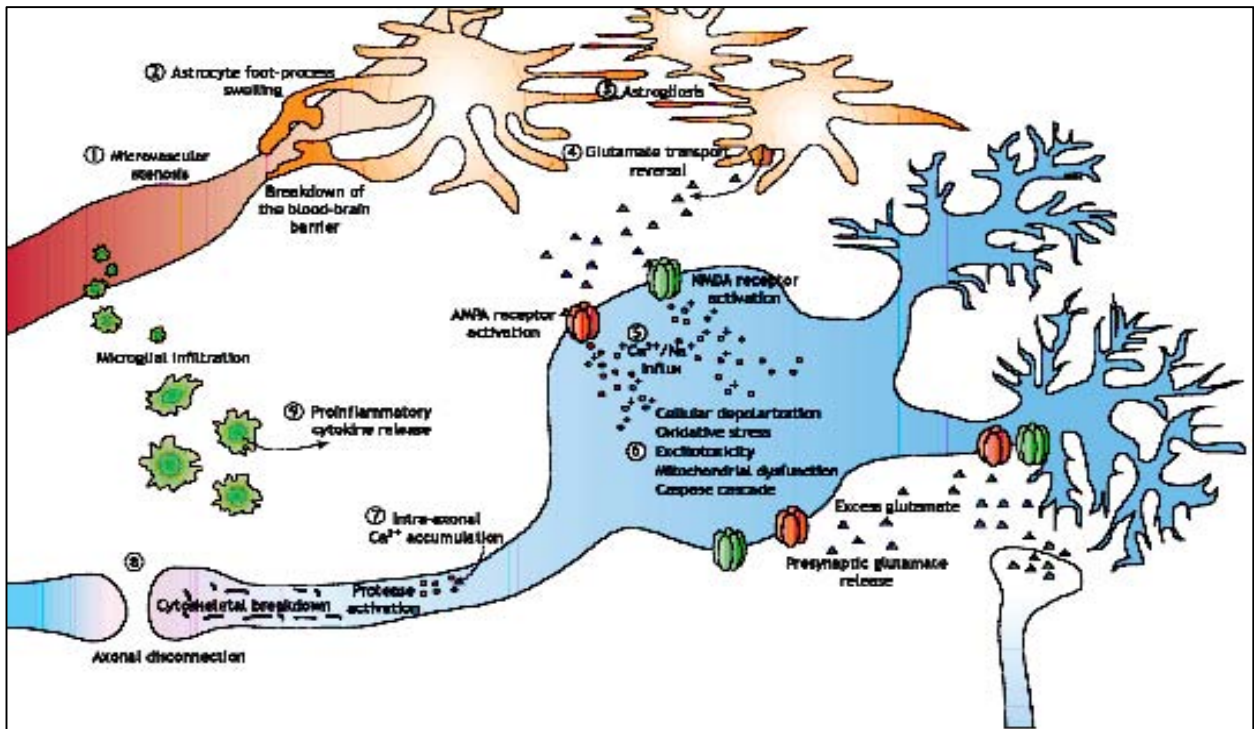
 , μ (protease
 calpain) [28, 29]. μ - μ μ
 . μ -
 [30, 31].

μ μ ,
 μ μ - μ
 μ . , μ -
 μ μ
 [32-35].
 μ NGF (nerve growth factor)
 μ μ [36].
 μ μ (Bcl-2)
 () μ
 μ μ
 [37].

μ
 μ , μ -
 (vasogenic edema) (cytotoxic edema).To μ
 μ , μ μ
 . μ
 μ μ μ μ
 μ ,

[38]. 45min 2
 [39]. VEGF (vascular endothelial growth factor)
 35% (<90 mmHg) (PaO2<60 mmHg).
 39% [40].
 [41]. (>60 mmHg)

[42].



:
 μ
 μ (1)
 μ
 μ
 μ (“ ”) (3)
 ,
 μ
 μ (excitotoxic) μ μ .
 , Ca (5) μ μ
 μ
 Ca Zn μ
 (6), AMPA NMDA
 μ
 μ
 μ
 μ Ca, μ
 μ
 μ (7) μ μ μ
 μ (8). μ μ μ
 , μ (9)
 μ
 [43].

μ μ μ μ IL-1
 μ μ IL-1
 μ μ , μ μ
 μ μ ,
[82].
IL-1 μ μ
. ,
, μ μ / μ . , μ μ
, [83]
IL-1 μ .
IL-1 μ μ μ IL-1 μ IL-1
, μ ,
IL-1 μ μ μ μ μ -
 μ [84]. μ
 μ μ IL-1 IL-1 μ [85,
86]. IL-1 , IL-1 ,
, μ , μ [87].

<p>mRNA</p> <p>, μ</p> <p>, μ , μ ,</p> <p>μ , μ ,</p> <p>μ</p>	<p>Fan et al. [88](1995), Shohami et al(1997), Szafarski et al[89] (1995)</p> <p>Yan et al. [90] (1992)</p> <p>Woodroffe et al. [91] (1991)</p> <p>Wang et al. [92] (1997)</p> <p>Rostworowski et al. [93] (1997)</p> <p>Herx et al. [94] (2000)</p>
<p>, μ</p> <p>, μ ,</p> <p>μ , μ ,</p> <p>μ , μ</p> <p>μ</p>	<p>Taupin et al (1993)</p> <p>Tchelingerian et al. (1993)</p> <p>Fagan and Gage [95] (1990)</p> <p>Yan et al. (1992)</p> <p>Wang et al. (1997)</p>

: **IL-1** μ μ

IL-1 , IL-1RA IL-1 μ

μ .

mRNA IL-1 μ mRNA IL-1 . ,

μ , mRNA IL-1RA 6h μ μ 5 μ

[96]. IL-1RA μ .

mRNA IL-1 μ .

IL-1 (IL-1RI) μ μ

(IL-1RII) 3h μ

[97].

-6 (IL-6)

μ IL-6 6-8h μ [61].

μ μ μ , μ

μ IL-6 μ μ , μ

μ μ [98].

μ μ IL-6

(IL-6R) μ μ -

IL-6 μ μ -

/ . , μ -

μ NGF

(nerve growth factor) [99]. , IL-6

μ μ .

IL-6 μ μ , μ ,

μ μ μ

IL-6 (IL-6R) [90, 100]. ,

$\mu\mu$ μ
 -1
 μ $IL-1$ $\mu\mu$
 μ $[101].$ $IL-1$ μ
 μ $[84, 102].$ $, in vivo,$ μ $IL-1$
 $[103-105].$ μ $IL-1$ $[103]$
 $, ICE (IL-1$ $converting enzyme) [106]$ μ
 $IL-1$ μ μ μ $, \mu$
 $[37].$ $, Knockout$ $IL-1$ $IL-1$
 μ
 μ $.$ μ $[107].$
 $IL-1RA$
 μ $IL-1RA \mu$
 μ $[108, 109].$
 μ $IL-1RA [110]$ $-$
 $[111-113].$ μ $IL-1$
 $IL-1RA$ μ μ
 $[114].$ μ $IL-1RA$ μ μ
 $[112].$ μ μ $, \mu$ $(fluid percussion injury)$
 μ $[109].$ $, IL-1RA$ μ $[115].$
 μ $IL-1RA$ μ
 μ $[116].$ $IL-1RA$

-6

μ IL-6
[117] μ μ μ , μ μ μ μ μ , Knockout
 μ μ μ μ μ μ μ μ
IL-6 (wild type mice) [118].

-18 (IL-18)

μ IL-1 IL-18,
IFN- (Interferon-gamma) , μ μ μ
 μ μ [77, 119-121]. μ
 μ IL-18
[122, 123] μ , μ ,
 μ μ IL-18 μ -
 , IL-18BP (IL-18 binding protein). , μ IL-18BP
 μ μ μ . μ , μ
 μ IL-18 , μ
 μ μ μ [122].

-12 (IL-12)

IL-12 μ - μ ,
IFN- [124]. , μ
[125]. μ
 μ . , μ μ
IL-12 , μ [126], [127],
[128]. , IL-12
 μ 14 μ μ μ , μ
 μ μ μ .

, - μ IL-1 μ .

, μ IL-1 μμ μ

μ (IL-4, IL-10 [129, 130] IL-1RA) μ μ

μ μ IL-6 (-

μ μ IL-1), μ

μ - μ .

μ , μ μ

IL-6 μ [131, 132]. ,

IL-6 [133]. IL-6 μ .

IL-1 μ

IL-1 IL-1RA μ

μ μ μ , μ

IL-1 μ μ μ

μ IL-1 (μ

) μ , IL-1RA [116] μ

μ , μ μ

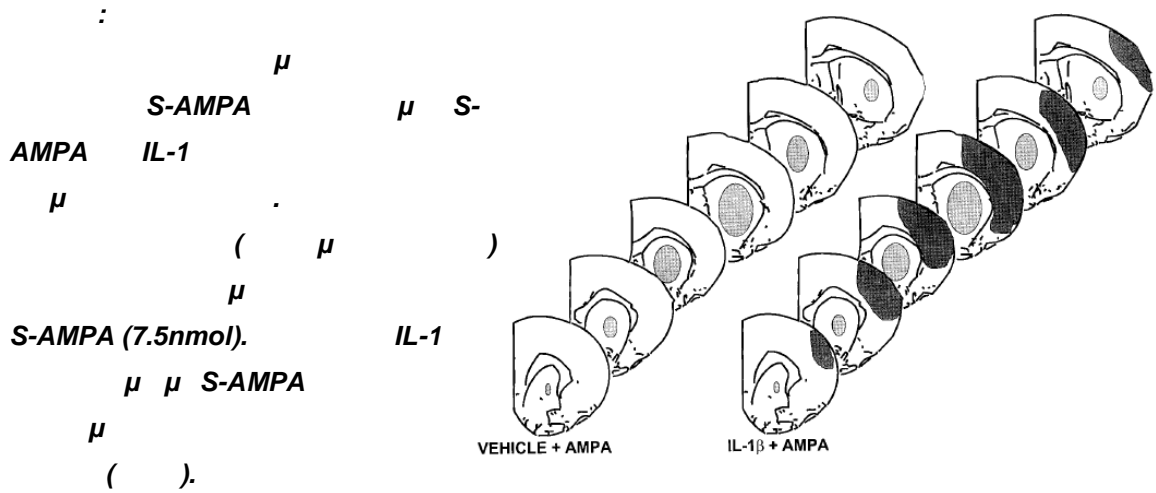
IL-1 μ μ

S-AMPA (-amino-3-hydroxy-5-methyl-4-isoxazole propionic acid: μ μ AMPA/kainate)

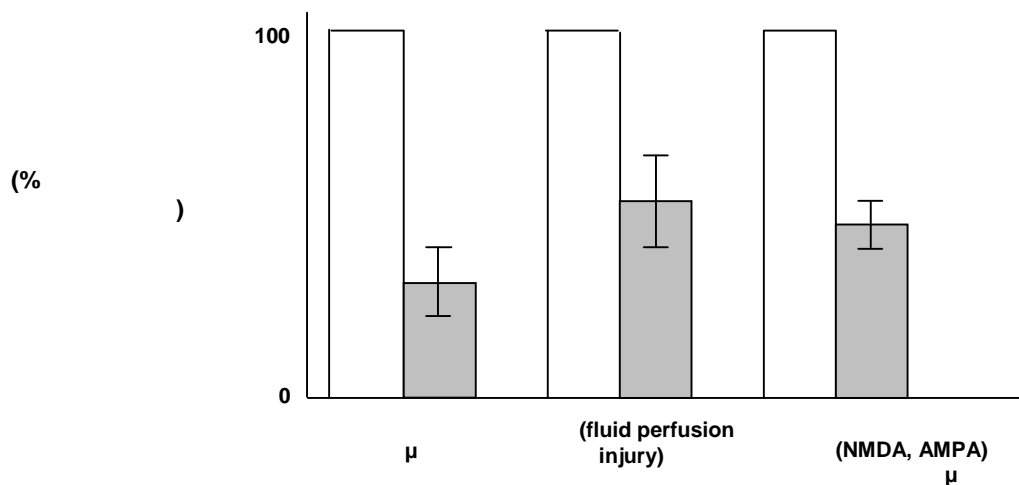
μ μ .

, IL-1 -

.



μ IL-1 IL-1RA
 μ ()
 μ ()
 μ ()
 μ ()
 μ ()



μ IL-1RA
 μ ()
 (lateral fluid perfusion injury, - 10 μ g IL-1RA),
 (5 μ g IL-1RA - 10 μ g IL-1RA),
 () NMDA AMPA μ

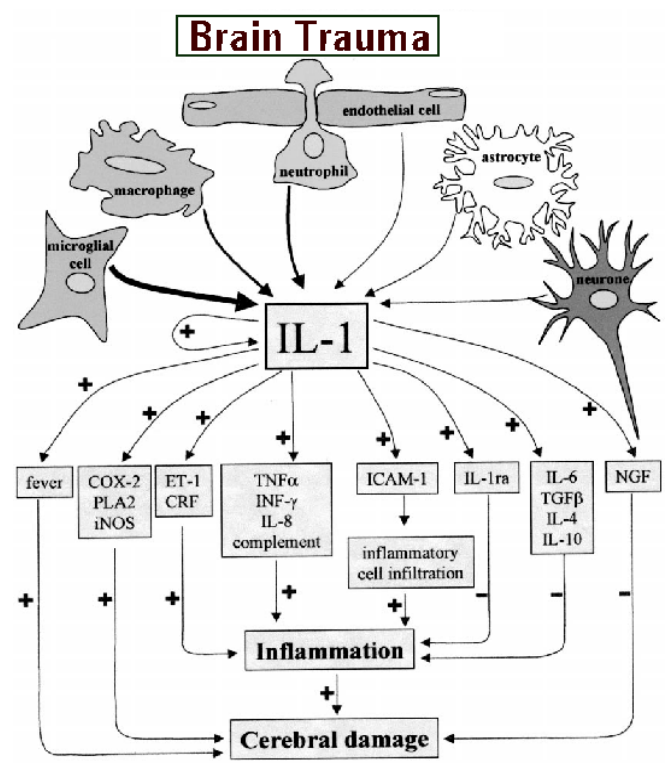
μ μ IL-1
 μ μ
 μ IL-1 μ μ
 μ μ μ
 μ μ
 μ μ μ
 μ μ μ
 μ μ μ
 μ μ μ
 μ μ μ

[134].

μ
 μ (. . . ICAM-1). μ
IL-1 μ [135]. μ , IL-1
 μ μ ,
[136].
: (1) μ mRNA IL-1
 μ μ [96], (2)
 μ ICAM-1
ICAM-1 μ μ μ [137,
138], (3) IL-1RA μ IL-1
 μ ICAM-1, μ
 μ μ [108, 135, 139]. μ , μ
 μ μ , μ IL-1
 μ μ .
, IL-1 ,
 μ , - μ
(IL-6, TNF , IFN- , IL-8, IL-1) - μ
(TGF- , IL-10, IL-4, IL-1RA) [140]. , μ
 μ μ μ ,
2.
, μ μ μ μ
IL-1 . , IL-1 in
vivo , IL-1 IL-1RA
in vivo in vitro μ .

μ , IL-1
 NGF (nerve growth factor)
 [141], μ IL-1RA [110]. μ ,
 IL-1 [142] [143] μ
 μ μ $\mu\mu$
 [78, 144]. μ μ o
 μ [145-147]. , -
 $\mu\mu$ μ -
 [148] μ [149, 150]. μ μ
 [151] μ
 NO NMDA [152].
 μ , μ IL-1
 μ IL-1
 μ

μ IL-1
 μ ,
 μ
 . COX-2: cyclo-
 oxygenase 2, PLA2: phospholipase 2,
 iNOS: inducible NO synthase, ET-1:
 endothelin 1, CRF: corticotropin
 releasing factor, TNF α : tumor necrosis
 factor, INF γ : interferon gamma, ICAM-
 1: intercellular cell adhesion molecule
 1, IL-1RA: interleukin-1 Receptor
 Antagonist, IL-6: interleukin-6, TGF :
 transforming growth factor, IL-4:
 interleukin-4, IL-10: interleukin-10,
 NGF: nervous growth factor.



IL-1 μ
 IL-1 μ
 μ [153]. , μ
 μ μ
 IL-1

IL-1 (5ng) μ μ -
 μ μ [104].
 IL-1 μ (2.5ng)
 μ [154].
 IL-1 μ -
 μ IL-1
 μ μ -
 μ IL-1RA -
 μ ICE μ μ μ ,
 [111, 115, 155]. μ IL-1RA
 μ ICE μ μ .
IL-1 μ
 IL-1 μ (CBF:cerebral
 blood flow) μ -
 μ μ [156]. ,
 IL-1 μ μ
 μ [97]. μ μ
 IL-1 ()
 μ , [157,
 158]. , IL-1 ([159]. μ , IL-1
)
 μ μ μ
 , μ μ
 μ μ IL-1RA μ μ
 μ ICE. μ IL-1RA
 μ ICE
 μ μ [160].

IL-1

IL-1 μ Ca^{2+} [161],
[162], μ iNOs (inducible nitric oxide
synthase) NO [163],
[164] CRF (corticotropin releasing factor) [165],
 μ μ - μ [166, 167],
 μ [168]. , IL-1
Ca²⁺, GABA ,
· μ , μ IL-1
· , IL-1
 μ μ μ
 μ [78].

IL-1

μ
IL-1 μ
Parkinson, μ Down , ,
IL-1 μ Alzheimer.
 μ , IL-1 , μ -
 μ [55, 169].
 μ μ Alzheimer (AD) IL-1
APP [170] (amyloid precursor protein) -
 μ μ - μ (A) μ μ μ -
· , IL-1
tau [171] () $\mu\mu$
 μ .
 μ μ μ -
IL-1 (IL-1 IL-1) μ
Alzheimer [172, 173]. , μ μ μ μ
IL-1 μ AD.
 , μ

IL-1. IL-1

IL-1

IL-1

Alzheimer (AD) [174].

IL-1 [175]

[169]. IL-1

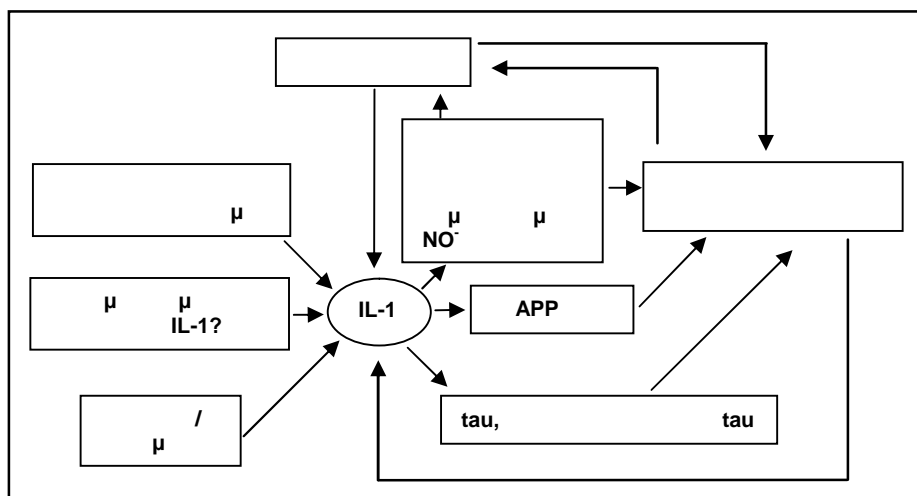
AD.

AD

[176]

APP tau [177]

IL-1RA APP

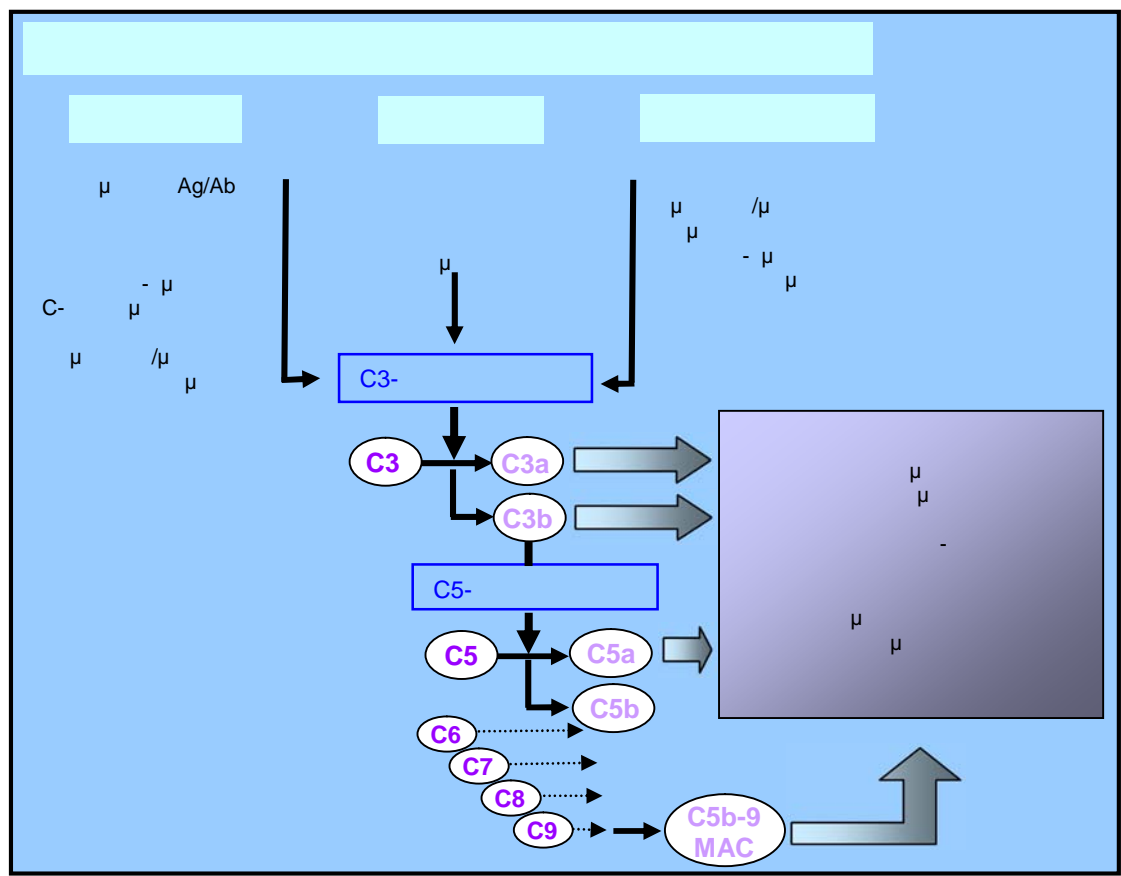


IL-1 IL-1 apoE

IL-6 μ

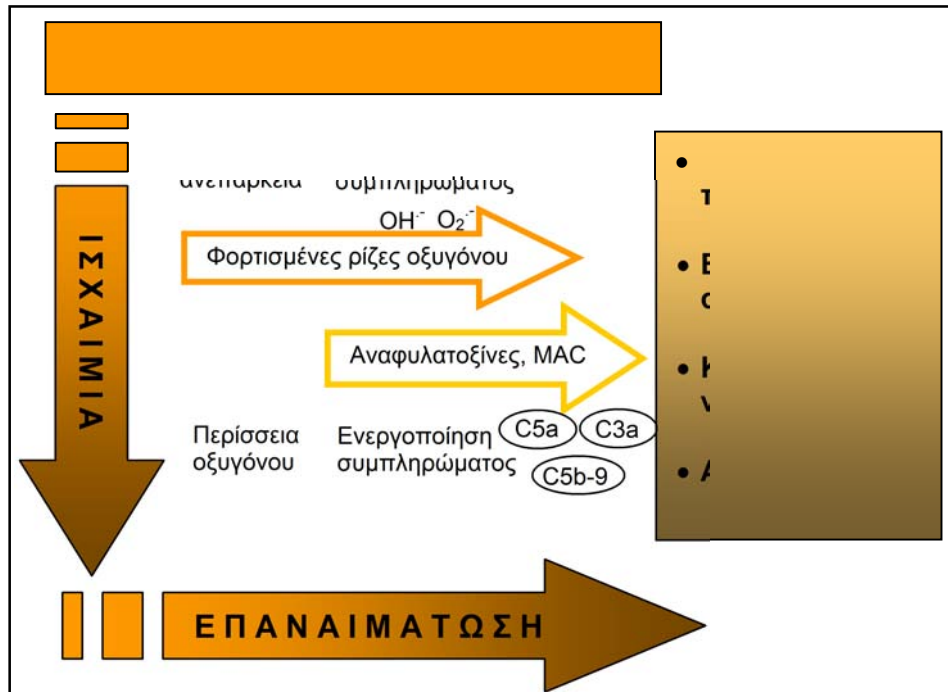
, μ μ -
IL-6 μ μ
IL-6
 μ ,
 μ
 μ μ μ μ -
 μ , IL-6 μ
[178].
 μ (μ IL-1 , TNF-) [179, 180] -
 μ - (HPA axis: hypothalamic-pituitary-adrenal axis), μ -
 μ ,
IL-6 μ -
 μ , μ ,
necrosis factor, μ), TNF (tumor
growth factor), NGF (nerve
NMDA (N-methyl-D-
aspartate) [57]. , IL-6
 μ
 μ μ μ
 μ , μ μ .
 μ μ μ , μ -
 μ , μ , μ -
 μ μ μ [181, 182]. μ -
 ,

184]. [183, -
 [185-188]. [190, 191],
 [189]. C3a C5a),
 C5a receptor-C5aR
 (MAC: membrane
 attack complex/C5b-9),
 [192-194].
 [58, 195].



: μ μ μ μ μ
 μ
 . MAC: membrane attack complex. C3 μ C3
 μ μ C3b μ ,

μ μ μ , - . μ
 μ μ μ μ
 μ μ [200, 202]. , μ
 μ
 μ [203].



7: μ / μ
 μ . μ μ μ
 μ μ . μ
 (nutritive failure) μ
 (ATP: adenosine triphosphate), μ μ
 . μ , μ
 μ , μ μ μ
 μ μ , C3a C5a, μ MAC
 (membrane attack complex)/C5b-9. , μ / μ
 μ μ

IL-1

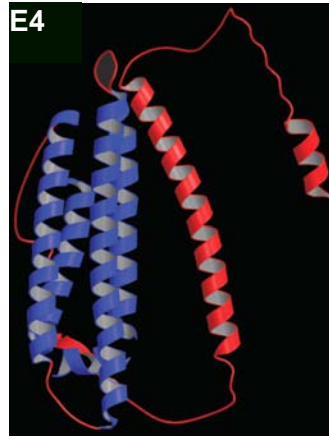
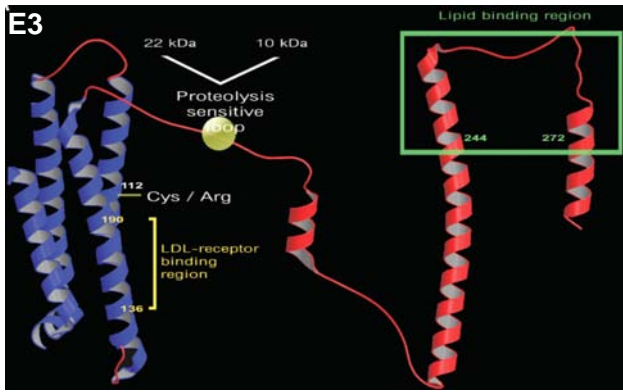
IL-1 is a pro-inflammatory cytokine that plays a central role in the initiation and regulation of the inflammatory response. It is produced by various cell types, including macrophages, monocytes, and T cells, in response to various stimuli such as lipopolysaccharide (LPS), endotoxins, and other pathogens. IL-1 is a dimeric protein consisting of two identical subunits, IL-1 α and IL-1 β , which are linked by a disulfide bond. IL-1 β is the biologically active form of the cytokine, and its production is regulated by the transcription factor NF- κ B. IL-1 acts on target cells through its receptor, IL-1R, which is a heterotrimeric complex composed of IL-1R1, IL-1RAcP, and a signaling subunit. IL-1R1 is a type I transmembrane receptor with an extracellular domain that binds to IL-1 β . The binding of IL-1 β to IL-1R1 leads to the recruitment of IL-1RAcP and the signaling subunit, which then activates the MyD88-dependent signaling pathway. This pathway leads to the activation of NF- κ B, which in turn induces the expression of various pro-inflammatory genes, including IL-1, IL-6, TNF- α , and IL-8. IL-1 also plays a role in the regulation of the immune response, including the activation of T cells and the production of other cytokines. IL-1 is involved in the pathogenesis of various inflammatory diseases, including rheumatoid arthritis, Alzheimer's disease, and atherosclerosis. IL-1 is also involved in the regulation of the circadian rhythm and the sleep-wake cycle. IL-1 is a key player in the inflammatory response and is involved in the pathogenesis of various inflammatory diseases.

μ μ μ ,
 μ μ
 μ . μ
 μ
 μ , , COMT
 (catechol-o-methyltransferase), neprilysin, DRD2 (dopamine
 D2 receptor), , p53 , ACE
 CACNA1A μ .

(ApoE)

apoE

APOE μ 3.7Kb, 4
 μ μ μ 19 [209]. apo μ
 (=34KDa) 299 μ μ
 μ μ μ [210].
 3 μ apoE: 2, 3 4 3
 : 2, 3 4. μ
 (cys) (arg) 112 158
 μ , 2 cys¹¹²cys¹⁵⁸, 3
 cys¹¹²arg¹⁵⁸ 4 arg¹¹²arg¹⁵⁸. μ apoE μ -
 μ 7% (2), 78% (3) 15% (4) μ [211].
 μ μ APOE :
 3 μ (APOE 2/2, APOE 3/3, APOE 4/4) 3
 (APOE 2/3, APOE 3/4, APOE 2/4). μ
 APOE 2/3, APOE 3/3, APOE 3/4.
 4
 Alzheimer (AD) [212-214], μ μ
 μ [215] μ [216],
 μ μ μ
 [217, 218].



: μ 3 4 μ apoE

apoE

, apoE μ .
 μ , μ μ .
 μ , μ μ - , μ -
 apoE, μ μ μ .
 μ apoE μ μ μ μ ,
 μ [219], μ ,
 apoE μ μ , apo LDL (low
 density lipoprotein) μ μ μ μ
 , apo LDL, μ μ apoE
 μ .

apoE

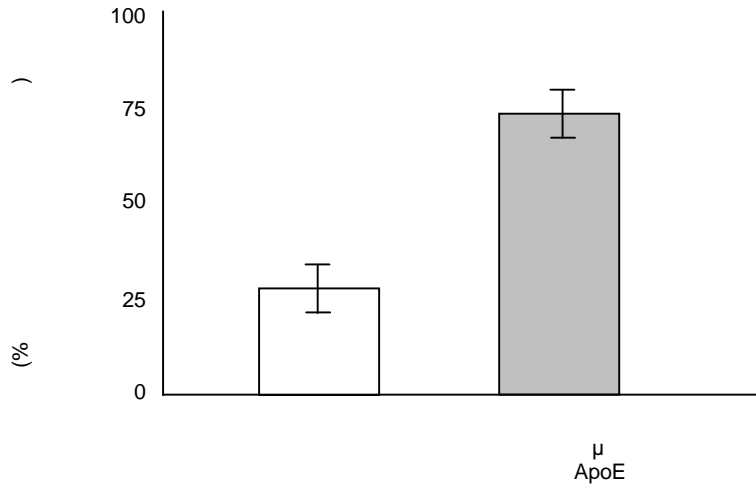
apoE μ ,
 μ μ μ
 μ [220]. μ μ
 apoE μ μ
 μ . μ μ APOE

[221],

-

apoE μ

[222].



:

μ

μ

μ

APOE

μ

,

μ

4

μ

-

μ

μ

μ

.

μ

μ

apoE

μ

-

μ

μ

μ

μ

AD [223].

30%

μ

μ

[224].

μ

μ

μ

apoE

μμ

μ

AD

apoE

AD.

μ

μ

APOE

4

4

μ

μ

4

[225].

μ

μ APOE 4

μ

μ

μ

[218]

μ

(n=93)

57%

μ 4

μ

(

,

,

μ

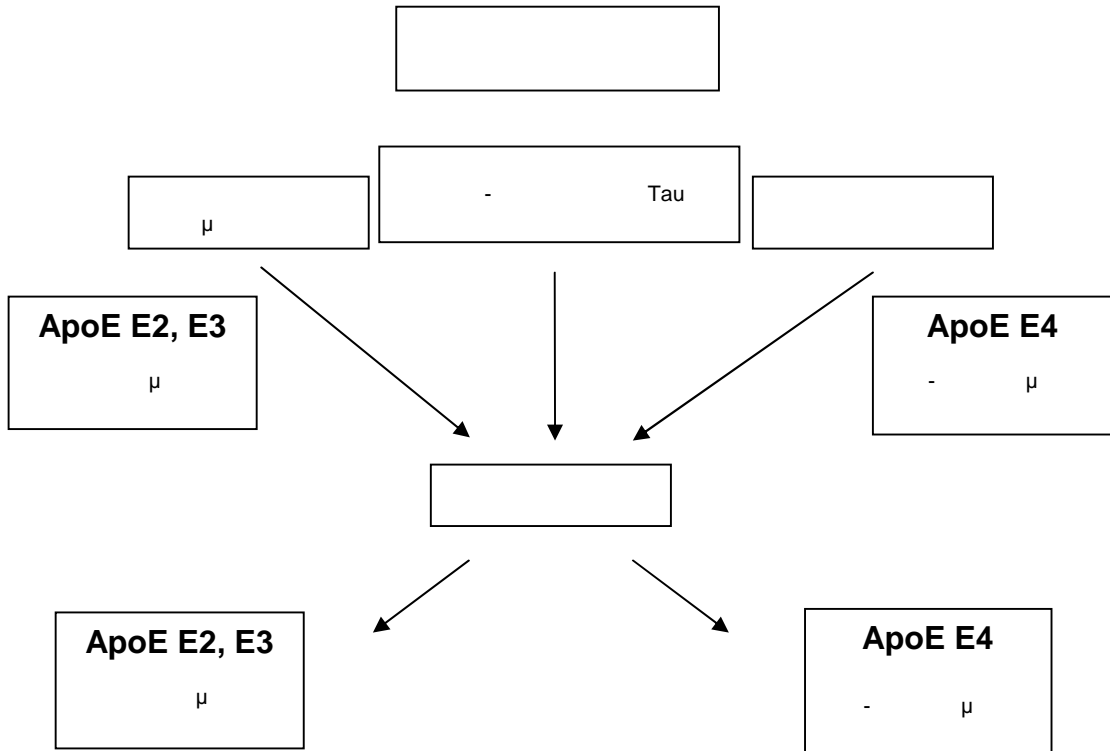
μ

) 6 μ μ μ 27%
 4 μ . μ
 μ ,
 (μ μ , [GCS]) μ
 μ . , μ 4
 (μ GCS) μ .
 μ μ μ
 μ [226]. μ μ
 μ μ apoE μ μ μ μ
 μ μ , μ μ
 μ 4 [227].
 , APOE
 μ μ μ [217]
 4 μ -
 μ μ μ (>7 μ).
 , μ μ μ μ -
 4 μ -
 [228]. , 4
 μ μ μ μ [229],
 μ μ μ [230].
 μ APOE -
 μ μ . μ ,
 4 μ
 μ μ μ μ -
 [231]. μ , μ μ [232] μ
 μ 4
 μ μ μ μ - . ,
 4 μ
 μ μ μ μ [233] μ μ - .
 , μ μ 77 μ
 4 μ
 μ μ μ μ -
 [234]. μ μ -
 μ APOE
 μ , μ
 μ [235].
 APOE μ μ ,

apoE

1. apoE [236]

APOE [237].



APOE

2. apoE

(lipid peroxidation),

[238].

3. apoE
 μ μ [239] μ
 [240].

4. apoE
 μ μ
 3 μ μ μ μ
 μ tau (μ) μ -
 μ μ , 4 μ
 μ tau [241].

PO

APOE

μ μ . μ ,
 G/T μ μ -219 A/T μ μ -491 .
 μ -219 -
 169% . , -
 μ -491 μ 63%
 [242]. μ , -
 μ AD. μ ,
 G-219T μ μ -491 μ -
 μ AD [243]
 μ μ AD [244, 245]. μ
 μ μ [246] μ
 μ μ ,
 G-219T μ μ
 μ GG GT . μ -491 /
 μ μ .

Neprilysin

- μ μ μ
 μ μ μ
 μ . μ , μ
 Alzheimer

neprilysin [247]. AD
 neprilysin
 [248, 249]. knockout
 neprilysin
 [247]. neprilysin
 [250]. neprilysin
 [251].
 GT neprilysin
 AD [252, 253].
 neprilysin
 DNA,
 81
 3.45
 GT [254].
 neprilysin

COMT

COMT (catechol-o-methyltransferase),
 (DA) (NE),
 : (COMT Val/Val, COMT Val/Met, COMT Met/Met).
 DA [255],
 DA [256, 257]. [258] COMT
 Val158Met
 (Val/Val), DA,
 (Wisconsin Card Sorting Test, WCST),
 (Met/Met) DA
 39 27

μ Val μ μ ,
μ μ (Continuous Performance Test).

Dopamine D2 receptor

DRD2 (dopamine D2 receptor gene)

μ μ DA, ,
C/T μ μ
Taq1 DRD2 . μ μ
μ (C/C, C/T, T/T). μ (Taq1 A)

μ μ 40% D2
[259]. , μ
AD [260]. , μ

μ μ [261] μ μ μ 39
μ 27 μ μ μ
μ

CVLT (California Verbal Learning Test)

μ μ - . , CPT μ
(Continuous Performance Test, μ μ)

μ μ - μ . , μ μ

μ μ μ μ μ

μ μ - μ μ μ -

μ μ μ μ μ

μ μ μ μ μ

μ μ μ μ μ

-1 (IL-1). μ μ :

IL-1 , IL-1 , IL-1 Receptor Antagonist (IL-1RA). IL-1 IL-1 -
 μ IL-1A IL-1B ,
 μ μ μ
2 [262]. IL-1RA IL-1 IL-1 ,
IL1RN [263]. μ μ 5
 μ IL-1A , -889 [262]. μ IL-
1A*2 μ μ μ μ AD [172, 173,
264, 265]. IL-1B μ .
 μ μ 5 IL-1B , +3953.
 μ 2 μ μ
IL-1 μ μ 1
[266]. μ μ IL-1B
, -511 [267]. , IL1RN
 μ μ 2
 μ (variable number of tandem repeats). μ μ
5 , μ IL-1R 2 μ μ
IL-1 in vivo. , μ μ IL-1B +3953,
IL-1B-511 IL-1RA μ μ μ AD [173, 268].
 μ μ μ
. μ μ [269] μ μ μ
IL-1B μ . μ , 14
25 (56%) IL-1B(+3953)*2 μ -
 μ 8 44 (18.1%) μ - 2. , 20
28 (71.4%) IL-1B(-511)*2 μ -
 μ 2% (2 41) μ - 2 (P=0.005).
, IL-1A (-889) μ μ μ μ
[270].
-6 (IL-6) μ μ -
 μ μ .
 μ IL-6 μ μ
 μ [271, 272]. , μ μ
IL-6 (-174) μ μ ,
 μ [272].

p53

[37]. p53
 [273] μ (pro) (arg) μ 72 -
 [274]. μ μ
 , Arg/Arg μ
 [275]. μ μ [276] Arg72Pro
 μ μ μ μ . μ
 μ μ - , Arg/Arg -
 (69%) μ Arg/Pro Pro/Pro (31%).
 , Arg/Arg μ μ (2.9)
 μ - μ μ p53 μ .

Angiotensin converting enzyme (ACE)

μ μ (ACE)
 μ - , μ μ .
 ACE μ μ ,
 [277]. ACE μ
 μ μ 17 17q23.
 μ μ / (I/D: insertion/deletion)
 16 ACE (II, ID DD),
 ACE μ [278]. μ ,
 ACE μ μ μ DD
 [279]. , I μ μ
 μ AD [280, 281], D μ μ -
 μ [282-284] μ μ
 74 [285].
 μ μ [286] μ ACE I/D μ μ
 μ KEK.
 D μ μ
 μ (Grooved Pegboard Trial
 Making Tests) μ I . μ μ

		μ μ	μ μ μ	μ	μ
Nicoll et al (1995)[225]	APOE	2/ 3/ 4	90 (23 A +)	μ	APOE4 , p<0.00001
Sorbi et al (1995)[228]	APOE	2/ 3/ 4	16	μ μ	APOE4 , p<0.01
Mayeux et al (1995)[174]	APOE	2/ 3/ 4	236	μ μ μ	μ AD APOE4 μ AD μ
Katzman et al (1996)[289]	APOE	2/ 3/ 4	160 μ μ AD, 69	μ AD	APOE4 μ AD, OR 13.5, 95% CI 2.63-69.12, p=0.0018
Teasdale et al (1997)[218]	APOE	2/ 3/ 4	89	μ	APOE4 , OR 0.23, 95% CI 0.06-0.82, p=0.024
Jordan et al (1997)[227]	APOE	2/ 3/ 4	30 μ	(CBI μ)	μ APOE4 p<0.01
Friedman et al (1999)[217]	APOE	2/ 3/ 4	69	μ	APOE4
Mehta et al (1999)[290]	APOE	2/ 3/ 4	797	μ μ 2.1	μ AD μ μ
Plassman et al (2000)[291]	APOE	2/ 3/ 4	46	μ AD, 356 AD, μ	μ AD
Lichtman et al (2000)[292]	APOE	2/ 3/ 4	31	μ μ μ 6 μ	μ μ μ μ APOE4 μ μ , p=0.05
Guo et al (2000)[293]	APOE	2/ 3/ 4	942 327	μ AD μ	μ AD APOE4 , OR 3.3, 95%CI 2.0-5.5
Kutner et al (2000)[294]	APOE	2/ 3/ 4	53	μ	APOE4 μ μ μ p=0.004
Liaquat et al (2002)[229]	APOE	2/ 3/ 4	129	μ μ μ μ GOS μ 6 μ	E4 μ μ μ μ APOE4 , p=0.0056 , p=0.015
Crawford et al (2002)[233]	APOE	2/ 3/ 4	110	μ μ μ μ 6 μ μ	APOE4 μ μ μ μ
Lieberman et al (2002)[231]	APOE	2/ 3/ 4	87	μ μ μ 3 6	APOE4 μ μ μ μ μ
Chiang et al (2003)[226]	APOE	2/ 3/ 4	100	μ μ GOS μ 6 μ	APOE4 , OR 3.01, 95%CI 1.02-8.88, p=0.04
Nathoo et al (2003)[235]	APOE	2/ 3/ 4	110	μ μ μ GOS μ 6 μ	-

Diaz-Arrastia et al (2003)[230]	APOE	2/ 3/ 4	106	μ μ	μ GOS μ 6 μ	μ E4 , OR 2.41,95%CI 1.15-5.07, p=0.03
Millar et al (2003)[295]	APOE	2/ 3/ 4	396	μ	μ μ 6 μ , 18	-
Kerr et al (2003)[296]	APOE	2/ 3/ 4	91	μ	μ μ () () , μ / ()	μ μ APOE4 / μ
Chamelian et al (2004)[297]	APOE	2/ 3/ 4	90	μ μ	GOS,	-
Sundstrom et al (2004)[232]	APOE	2/ 3/ 4	34	μ μ	μ	APOE4 μ μ μ μ μ
Quinn et al (2004)[199]	APOE	2/ 3/ 4	106	μ 2-19	μ	-
Koponen et al (2004)[298]	APOE	2/ 3/ 4	60	μ	30 μ μ	APOE4 μ , p=0.028
Teasdale et al (2005)[299]	APOE	2/ 3/ 4	1094	μ	GOS 6 μ	E4 μ μ - μ <15 , OR 3.06, 95%CI 1.22- 7.65, p=0.07
Leclercq et al (2005)[300]	APOE	2/ 3/ 4	88	μ μ	μ	E4 μ μ , p=0.021
Ariza et al (2006)[234]	APOE	2/ 3/ 4	77	μ	6 μ μ	APOE4
Smith et al (2006)[301]	APOE	2/ 3/ 4	239		μ	APOE4 , p=0.05 p=0.08
Jiang et al (2006)[302]	APOE	2/ 3/ 4	110	μ	(<7 μ μ)	APOE4 , OR 4.84, 95%CI 1.44-16.21, p=0.011
Isoniemi et al (2006)[303]	APOE	2/ 3/ 4	58	μ	μ μ , 31.3 μ	-
Kerr et al (2006)[296]	APOE	2/ 3/ 4	54	μ	Xe-CT μ μ μ μ 24h μ	APOE4 μ μ
Ponsford et al (2007)[304]	APOE	2/ 3/ 4	120	μ μ	6, 12 μ 3,	-
Han et al (2007)[305]	APOE	2/ 3/ 4	78	μ μ	1 μ	APOE4 μ
Alexander et al (2007)[9]	APOE	2/ 3/ 4	123	μ	GOS 3, 6, 12 24 μ	APOE4 μ

Hiekkanen et al (2007)[306]	APOE	2/ 3/ 4	33 μ μ -	MRI μ 1 μ 1	-
Zhou et al (2008)[307]	APOE	2/ 3/ 4	- : 14 μ μ 2427 μ μ	GCS μ GOS 6 μ	- APOE4, RR 1.36, 95%CI 1.04-1.78
Willemse-van Son et al (2008)[308]	APOE	2/ 3/ 4	79 μ μ	GOS 3, 6, 12, 18, 24, 36 μ μ	APOE4 , OR 0.26, 95% CI 0.02-0.51, p=0.037
Tanriverdi et al (2008)[309]	APOE	2/ 3/ 4	93 μ , 27 μ	μ	μ APOE3/E3 μ , OR 0.29, 95% CI 0.11-0.78, p=0.01
Brichtova et al (2008)[310]	APOE	2/ 3/ 4	70 μ	GCS , GOS 1	μ APOE4 μ - μ
Luukinen et al (2008)[311]	APOE	2/ 3/ 4	134 μ >70 , 28 μ μ ,	μ 9	μ ApoE4 , OR 2.70, 95% CI 1.02-7.16
Ost et al (2008)[312]	APOE	2/ 3/ 4	96 μ	μ 1 μ	- APOE4 μ μ μ (p=0.0079)
Rapoport et al (2008)[313]	APOE	2/ 3/ 4	49 μ , 68 μ	μ 1 2	-
Kristman et al (2008)[314]	APOE	2/ 3/ 4	318 (28 μ)		-
Ashman et al (2008)[87]	APOE	2/ 3/ 4	54 μ , 40 μ	2-5 μ ,	-
Han et al (2009)[315]	APOE	2/ 3/ 4	46 μ	μ	APOE4 μ μ
Muller et al (2009)[264]	APOE	2/ 3/ 4	59 μ	6 μ μ	ApoE4 μ μ , p=0.046
Hiekkanen et al (2009)[316]	APOE	2/ 3/ 4	33 μ	1 μ μ μ μ , GOS	-
Lo et al (2009)[317]	APOE	2/ 3/ 4	65 μ	μ (CPP)	μ μ μ CPP ApoE4 μ μ - , p=0.03
Lendon et al (2003)[246]	APOE	-219G/T -491A/T -427C/T	92 μ	GOS 6 μ	μ - μ μ
Jiang et al (2007)[318]	APOE	-219G/T -491A/T -427C/T	110 μ	(<7 μ μ)	-491AA μ APOE4
Jiang et al (2008)[319]	APOE	-219G/T -491A/T -427C/T	110 μ	(<7 μ μ CT)	- - -
Terrell et al (2008)[320]	APOE	2/ 3/ 4	195		-

	Tau APOE	His47Tyr Ser53Pro -219G/T	72 μ μ μ 8			- - TT , OR 2.7, 95% CI 1.1-6.8, p=0.03
Johnson et al. (2009)[254]	Nepriylisin	GT	81 μ μ μ μ	μ μ A		μ μ GT >41, OR 10.1, 95% CI 3.1-32.5, p=0.0001
Martinez et al (2005)[276]	p53	Arg72Pro	90 μ	μ GOS GOS μ 6 μ		Arg/Arg, OR 2.9, 95% CI 1.05-8.31, p=0.039
Ariza et al (2006)[286]	ACE	I/D	73 μ μ	μ μ μ μ		D μ
Uzan et al. (2005)[321]	IL-1b	+3953 C/T -511 A/G	69 μ	μ GOS 6 μ μ GOS 6 μ		Allele 2, OR 0.25, 95% CI 0.12-0.55, p= 0.0004 Allele 2, p=0.005
Hadjigeorgiou (2005)[322]	IL-1RA IL-1b	VNTR -511 A/G	151 μ	μ		Allele 2, OR 4.57, 95% CI 1.67-12.96, p = 0.004 -
Tanriverdi et al (2006)[270]	IL-1a	-889 C/T	71 μ	GOS 6 μ		-
Dardiotis et al. (2006)[323]	IL-1a	-889 C/T	215 μ	GOS 6 μ		-
Minambres et al (2003)[272]	IL-6	-174 C/G	62 μ	GOS 6 μ		-
McAllister et al (2005)[261]	ANKK1	rs1800497	39 27 μ μ	(SD=24.4) μ μ 38.4 μ		T μ μ μ
McAllister et al (2008)[261]	ANKK1 NCAM DRD2	31 polymorphisms	93 μ 21	(SD=15.8) μ μ 43.1 μ		T SNPs (rs1800497) μ ANKK1
Lipsky et al (2005)[258]	COMT	Val158Met	113 μ	μ		Val μ
Chan et al (2008)[324]	SLC6A4	5-HTTLPR rs25531	174 μ			-

-1 (IL-1)

IL-1 IL-1

IL-1 3 3
 : IL-1 IL-1 , IL-1RA (IL-1
 Receptor Antagonist) IL-1. IL-1 IL-1
 μ μ μ
 () μ μ 31KD. μ
 μ μ 17KD μ μ
 . , μ μ IL-1RA (secreted IL-
 1RA: sIL-1RA) μ μ
 μ .
 IL-1 (IL-1R): I (IL-1RI)
 μ , II (IL-1RII) -
 μ IL-1 μ μ μ .
 IL-1 I μ , μ
 AcP (IL-1R-accessory protein) [325]. μ μ μ -
 IL-1RI μ IL-1R-AcP μ -
 μ μ .
 IL-1RI (IL-1sRI) IL-1RII (IL-1sRII) μ IL-1 , IL-1
 IL-1RA.

IL-1

IL-1
 μ μ IL-1 μ
 μ μ 4.200 (bp)
 μ μ 200bp
 [326].
 IL-1
 μ IL-1 μ
 17KD. μ μ IL-1 (proIL-1) 31KD -
 μ μ μ (μ) , μ
 μ [327]. proIL-1

μ , μ μ proIL-1
 μ μ μ , -
 , proIL-1 -
 [177]. ,
 proIL-1 (calcium-
 dependent, membrane-associated cysteine proteases-calpains) [177, 328]. -

μ Ca^{2+} ,
 μ μ IL-1 . , μ
 IL-1 μ [329].

IL-1
 proIL-1 μ μ μ
 , IL-1 μ .
 IL-1 μ μ μ
 . μ
 μ [329].

μ IL-1
 In vitro μ proIL-1
 , μ - μ .
 10-15% IL-1 μ μ [330] μ
 [331]. μ μ (myristoylation)
 μ μ . μ μ IL-
 1 μ μ μ μ
 [332]. μ μ IL-1
 μ IL-1 (anti-IL-1) . , μ μ
 IL-1 (anti-IL-1) μ μ IL-
 1 .

IL-1

μ μ IL-1 IL-1
 μ μ μ
 [333, 334]. μ μ (μ

(μ) μ) μ [327, 335].
 μ μ μ μ
 [330]. , μ
 , mRNA, μ ,
 . , μ μ μ
 IL-1 μ IL-1 μ
 IL-1 . μ μ IL-1 μ
 μ μ IL.
 IL-1 , IL-1 μ -
 μ μ .

IL-1

μ IL-1
 μ μ mRNA IL-1
 15min,
 μ μ 4 .
 μ / μ μ mRNA [336,
 337]. IL-1
 mRNA IL-1 24 [239, 338].

, proIL-1 μ ,
 μ μ ICE (IL-1 converting enzyme).
 μ μ IL-1 , μ
 . μ μ IL-1 (aa 1-116) μ μ
 [330]. , μ μ μ proIL-1 ,
 μ μ μ IL-1
 [339]. IL-1
 μ [340] μ [327],
 μ μ μ IL-1 . , μ
 proIL-1 IL-1 μ μ μ
 μ 116-117 (asp-ala) ICE μ [15].

μ IL-1
 μ IL-1 ,
 , μ μ μ

mRNA (splicing),
 μ mRNA, 3 μ -μ μ .

ICE (IL-1 converting enzyme)

μ μ , [15]
 μ μ IL-1 116-117 μ -
 (asp-ala), μ μ - , μ μ IL-
 1 .

IL-1R (IL-1 Receptor)

μ IL-1:
 I (IL-1RI) II (IL-1RII).
 μ (lg) 3
 μ μ , μ μ [325, 341, 342].
 , IL-1RI IL-1RII
 μ μ 2 [343].

IL-1

IL-
 1 [344] 1984. μ
 μ IL-1 [345] IL-1 [346]. ,
 IL-1 μ 10 Kbp, IL-1
 7 Kbp. IL-1
 μ μ 2 (2q13) [347]. , IL-1 IL-1
 . ,
 μ
 μ [346].
 IL-1 IL-1 7 .
 μ 5 μ -μ μ mRNA
 . , μ μ
 IL-1. -
 IL-1
 [348]. , IL-

1 , IL-1 μ

μ μ (repetitive sequence elements), μ

Alu . IL-1 μ

IL-1

μ μ GRE (glucocorticoid response element)

(TGTYCT), ,

μ IL-1 . μ

3 μ -μ μ (3 UTR),

o - . 3 UTR

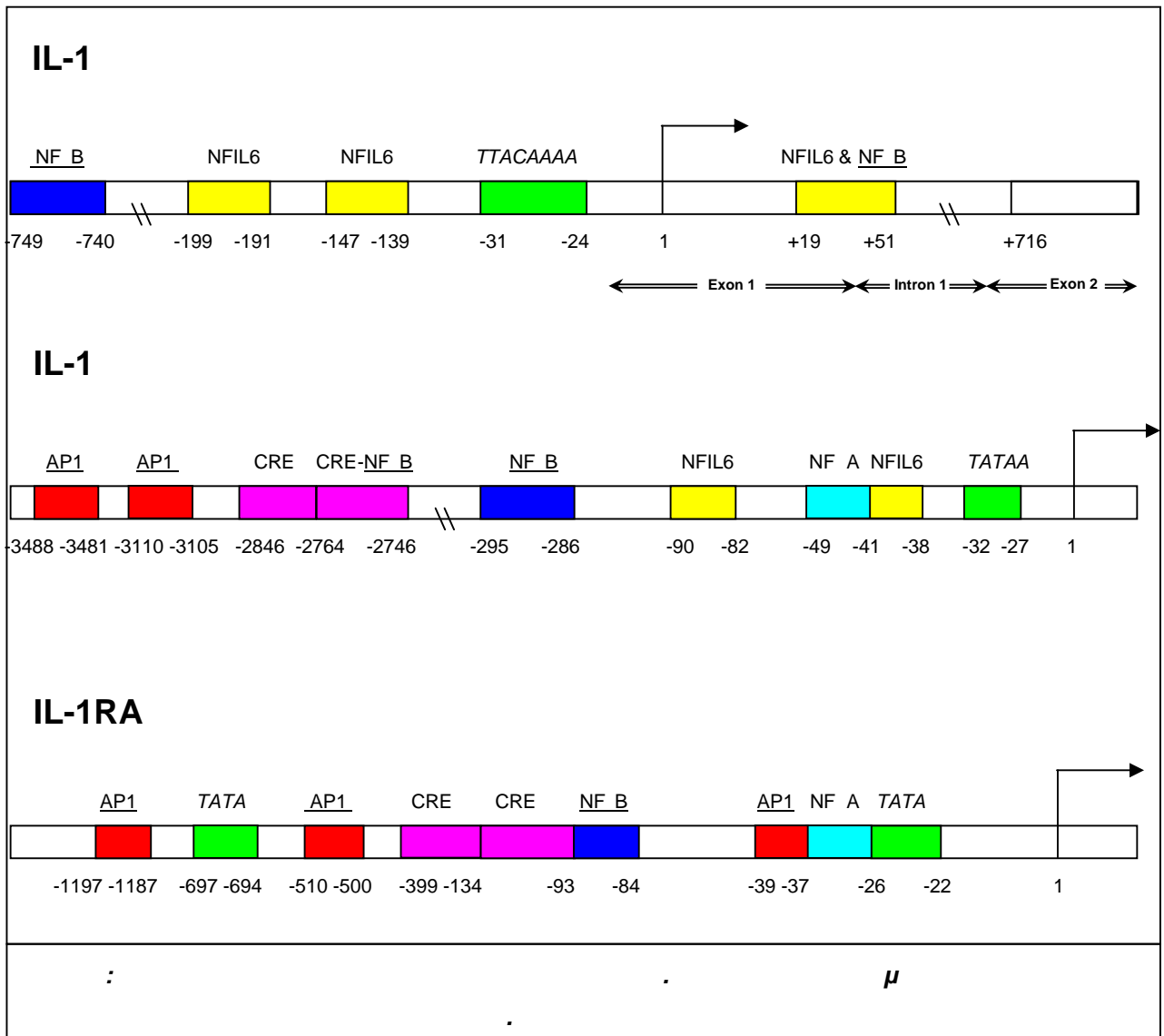
IL-1 IL-1 AU,

ATTTA μ .

μ ,

mRNAs - [349].

IL-1



motif), NFkB, IL-1RA, CREB (cAMP response element binding protein), [350].

IL-1, IL-1RA, NF-IL6, AP1, NFbA, TATA (TATA box IL-1RA).

(promoter) C/T polymorphism, SNP [267] (rs16944). IL-1 -511 (single nucleotide polymorphism, SNP) [351-353].

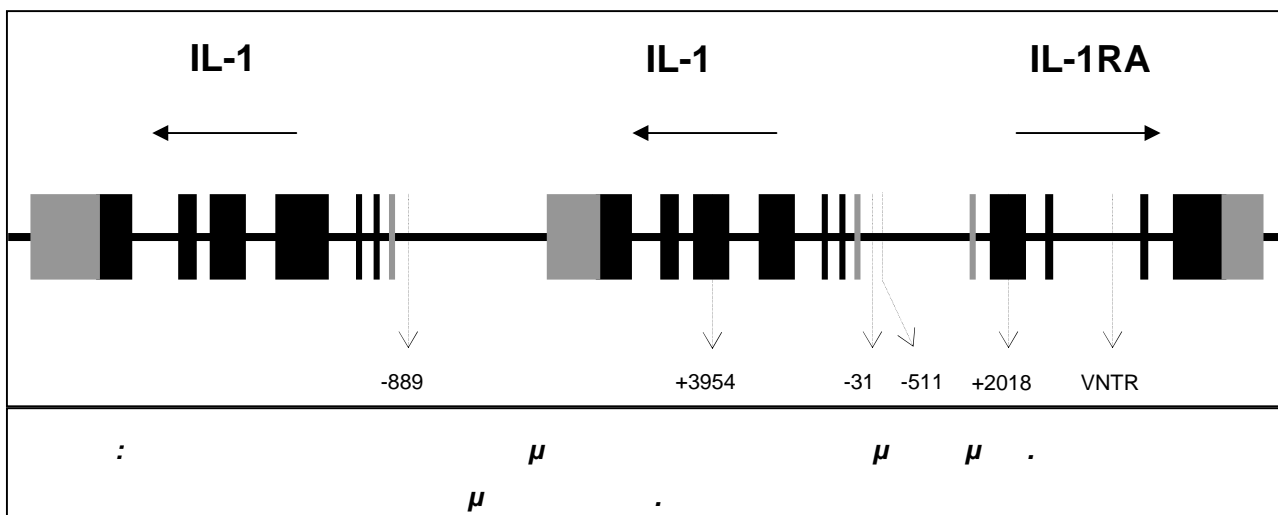
IL-1 .

IL-1B-511*1	✓	μ	μ	μ	[354]				
	✓				μ	[355]			
	✓					[356]			
	✓			Parkinson	[357]	μ	[358].		
	✓			[359]					
	✓			[360]					
	✓			μ	μ	μ	Alzheimer	[361]	
IL-1B-511*2	✓						[362].		
	✓			μ	μ		[363]		
	✓			μ	Sjogren	[364].			
	✓			μ		[365].			
	✓						μ	[366]	
	✓			Parkinson	[367, 368]				
	✓						μ		μ [369]
	:						μ	μ	IL-1 (-511 C/T) (rs16944)

IL-1 (-889) C/T polymorphism, SNP [370]. IL-1 (-889) IL-1 [371],

μ , $2/2 \mu$,
 μ IL-1 [372]. ,
 $\mu \mu \mu \mu \mu$. $\mu \mu$
 $\mu \mu \mu \mu$
 $\mu -92 -82,$
 -889 [373]. ,
 $\mu \mu$, $+4845,$
 pro-IL-1 IL-1 . $\mu -$
 μ IL-1 (-889) $\mu \mu$
 IL-1 (-889)*2 (IL-1 (+4845)*2) μ
 μ IL-1 [373].

IL-1 -889*1 ✓ $\mu \mu \mu$ [363] ✓ μ -Behcet [374] ✓ [375] ✓ [376] ✓ $\mu \mu$ [377]
IL-1 -889*2 ✓ μ Alzheimer [378] ✓ μ [379] ✓ μ [380] ✓ μ [381] ✓ μ [370]
: $\mu \mu$ IL-1 (-889 C/T)



IL-1

IL-1

μ μ μ μ μ in vivo in vitro

μ μ μ μ μ

μ μ μ IL-1 μ μ

μ μ Ca^{2+} [382], μ

μ [218], μ (long-term

potentiation) [383] GABA (- μ) [384].

, IL-1 in vivo μ

μ 2 (COX2) (inducible

nitric oxide synthase) (iNOS) [385]. , IL-1

μ CRF (corticotrophin releasing factor),

μ μ μ μ [154].

IL-1

IL-1

, μ [147]. ,

,

[386], (μ

- μ 1- μ) [59] μ μ [58].

IL-1 μ , -

, [387].

IL-1

μ μ μ (caspase) μ -

μ μ . -

IL-1 $\mu\mu$. μ ,

1 [160] (μ IL-1)

[155] μ μ

.

IL-1

IL-1 μ

μ IL-1 μ -

μ , μ [388] μ -

μ [84], [389]. ,

μ μ [390]

μ [388]. IL-1 μ

μ [391].

, IL-1RA , μ

μ , μ IL-1

, μ

μ [392]. μ μ

μ IL-1RA μ μ μ

apoE^{-/-} [393]. , μ μ knockout

μ IL-1 IL-1RA

apoE^{-/-} μ [394-398]. , μ IL-1RA^{+/-}

μ μ μ μ [396].

, μ μ μ

[399] knockout IL-1RA. μ

μ , μ

μ μ μ μ μ , μ -

μ μ μ μ .

μ ,

μ μ μ μ -

. μ μ μ -

μ μ

μ . , μ ,

μ - . μ

IL-1RA μ

IL-1, μ .

IL-1 μ

$\mu\mu$ μ

IL-1 μ

μ μ : μ μ [400]. IL-1

[401]

μ , μ μ

μ μ , IL-1 μ μ -

C [400]. μ C μ -

μ μ thrombomodulin. IL-1 -

,

C. , μ IL-1 $\mu\mu$

μ [402]. μ IL-1

μ μ [403].

IL-1 :

IL-1 in vivo

, in vitro μ IL-1

, μ

IL-1

μ NGF (nerve growth factor) [387]. μ , IL-1

[142] [143] μ μ -

μ $\mu\mu$ [144]. ,

μ IL-1

, , IL-1

μ .

IL-1 (IL-1RA)

IL-1RA 18% μ μ μ IL-1 26% μ

μ IL-1 . IL-1RA μ μ μ (sIL-

1RA) μ (icIL-1RA1,2,3) [404].

μ , sIL-1RA μ 17KDa,

177 μ μ (leader sequence),

μ .

sIL-1RA 4 3 .

, icIL-1RA μ 18KDa (icIL-

1RA1) 159 μ . μ IL-1RA

μ μ (splicing), $\mu\mu$
 icIL-1RA 9.4Kb upstream
 sIL-1RA. μ μ
 icIL-1RA sIL-1RA, 3
 μ (signal peptide). ,
 icIL-1RA μ , icIL-1RA
 μ μ .
 icIL-1RA μ , -
 μ μ μ .
 sIL-1RA IL-1, μ
 (icIL-1RA).
 , μ , ,
 , μ IL-1RA sIL-1RA
 μ mRNA
 μ IL-1RA, [405].
 μ IL-1RA
 [406]. μ μ μ 25KDa icIL-1RA2.
 mRNA icIL-1RA2 μ 63bp
 μ icIL-1RA.
 μ 3 N²⁻ μ μ icIL-1RA,
 μ μ μ 21 μ . mRNA icIL-1RA2
 μ , μ ,
 icIL-1RA2
 , mRNA μ μ in vivo.
 μ μ μ μ 16KDa icIL-1RA3 [407].
 μ μ , μ ,
 μ μ μ μ
 μ . , μ μ icIL-1RA,
 IL-1 μ μ IL-1RA.
 μ μ μ IL-1 μ
 μ μ sIL-1RA.
 μ IL-1RA -
 $\mu\mu$ -
 [404].
 IL-1RA μ ,
 , [408],

in vivo μ μ
 μ [404].
 IL-1RA μ
 μ μ ,
 μ (lg)
 3 μ μ . IL-1 -
 μ μ μ
 IL-1RA. IL-
 1 in vivo μ . ,
 IL-1RA μ μ
 μ IL-1.
 IL-1 IL-1RA μ -
 [404]; μ
 μ IL-1RA
 μ μ ,
 1 2 IL-1 IL-1 μ
 μ , IL-1RA μ . ,
 3 μ
 [409].

IL-1RA

IL-1RA (IL1RN) μ
 μ μ 2, q14-q21 [410]. IL-
 1 (IL-1), IL-1 (IL-1) IL-1RA , μ μ μ
 430Kb: IL-1 μ +0 +35Kb, IL-1 μ +70
 +110Kb IL1RN μ +330 +430Kb.
 μ
 μ μ 2,
 μ μ
 IL-1RA μ μ 2 (2-6)
 86bp [410]. 5 μ -

μ

μ μ .

atcctggggaaagtgagggaaatatggacatcacatggaacaacatccaggagactcaggcct
ctaggagtaactgggtagtgtgc

: **IL-1RN** **86** **μ** **5** **μ** **μ²** **VNTR**
μ μ (rs2234663).

μ

μ :

μ	μ	
1	4	μ
2	2	μ μ μ
3	3	
4	5	
5	6	

: **μ IL-1RA μ**

73%,

μ 2 μ μ 1 21%.

μ μ μ .

μ μ μ μ IL-1RA

μ μ IL1RN*2 μ μ μ μ μ

IL-1RA IL-1

IL1RN*2 μ μ IL-1 μ

μ μ IL-1 [351].

μ 2 μ μ

IL-1RA in vitro [411]. μ

sIL-1RA μ μ - . IL1RN*2 μ

μ μ icIL-1RA1 [404]

[412] μ 2 μ μ μ

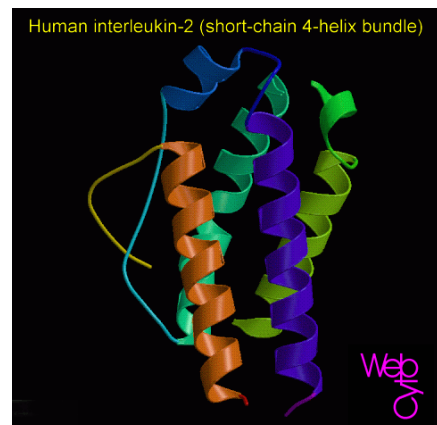
[404].

icIL-1RA1

2 (IL-2)

μ
 μ $\mu\mu$ μ μ
 μ Th1 Th2. Th1 μ CD4⁺T
 μ IL-2, IL-12 INF- Th2
 μ IL-4, IL-6
 IL-10. μ
 μ
 μ μ μ
 μ IFN-) μ Th1 (IL-2, IL-12,
 μ Th2 (IL-4, IL-6, IL-10)
 IL-2 μ μ 4 - (four-bundle -helical
 cytokine) μ - μ
 (IL-2R). IL-2 μ μ - μ
 μ μ μ μ μ μ μ -
 IL-2 63% μ
 μ

: μ IL-2



SHC (SRC-homology-2-domain-containing transforming protein C) STAT5 μ (signal transducer and activator of transcription) IL-2R .

MAPK (mitogen activated protein kinase) [419] PI3K-AKT μ (phosphatidylinositol 3-kinase) [420]. , tyr STAT5 μ -

‘ - ’ [421].

IL-2

IL-2 , μ

μ , μ μ

(T cell tolerance). IL-2

CD4+ CD8+ T μ μ ,

μ S

[422, 423]. , IL-2 μ

CD4+CD25+T μ ($_{reg}$) [424, 425],

(AICD: activation-induced cell death) [426]. , IL-2

μ NK (natural killer cells) ,

μ [427]. ,

IL-2 μ μ [428,

429].

IL-2 $\mu\mu$ μ ,

‘ ’ μ μ , μ , -

IL-2 μ . μ

IL-2 μ , μ -

μ (lymphoid hyperplasia) μ -

- μ , , μ

μ , μ [430] μ -

IL-2 μ CD4+CD25+T μ

($_{reg}$) μ (AICD),

μ - μ [426, 433]. μ ,
 IL-2 μ μ
 μ (peripheral tolerance).
 μ IL-2 μ -
 μ , μ
 μ (EAE-experimental autoimmune encephalomyelitis)
 [434], Crohn [435], μ [436],
 [437], μ μ (SLE-systemic
 lupus erythematosus) [438]. , μ μ -
 . , μ IL-2 μ μ μ
 μ , μ μ
 μ μ μ 1,
 μ (HTLV-I-human T cell leukemia virus type 1)[439],[383].

IL-2

IL-2 μ ,
 μ μ NK (natural killer cells) [440],
 - [441]. , IL-2 μ
 μ , μ [442].
 , IL-2 μ μ ,
 μ μ ,
 . , μ IL-2
 μ
 [443], μ μ
 μ .

μ IL-2

IL-2. IL-2 μ μ μ

μ [346] μ . μ
 μ , μ IL-2
 μ μ [444]. , μ
 EAE (experimental autoimmune encephalomyelitis)
 μ IL-2 (mRNA μ)
 μ μ
 μ [445, 446]. , μ μ
 IL-2
 () μ [346]. ,
 μ IL-2 μ ,
 μ ,
 [447, 448].

IL-2

IL-2 μ
 μ μ 4, q26-q27 [449]. μ IL-2,
 μ μ
 , μ μ μ
 , NF-AT (nuclear factor of activated T cells) , AP-1
 (activated protein-1), NF- κ B (nuclear factor κ B), -
 μ 1 (Oct-1)[450]. IL-2
 , μ
 NF-AT [451].
 μ μ μ μ
 NF-AT μ Jun Fos . μ μ
 c-Jun μ c-Jun/c-Fos
 NF-AT DNA. μ -
 μ μ IL-2,
 μ μ μ
 (TcR).
 PLC 1 μ (phospholipase C- -1) [452, 453],
 μ G- p21^{ras} [454, 455], μ
 μ NF-AT , μ
 μ IL-2 [450].
 IL-2 (IL-2R) 8 ,
 μ μ 25Kb μ μ 10. 2 4

IL-2R
 p15.1, IL-2R
 IL-2R
 q13 [457].

IL-2
 IL-2
 -384 IL-2 [458],
 (G) ().
 2. : G (), IL-
 () .
 -384 IL-2 (G/G)
 IL-2
 G/T T/T [361].

IL-2, +114 1 [459].
 : G (), T () .
 ,
 .
 IL-2 73%,
 G 27% [458, 460].

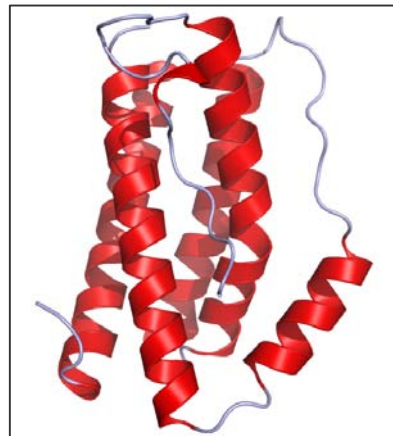
IL-2
 IL-2
 (T/G, T/T) -384
 IL-2
 [461].

μ
 μ (/) -384 μ μ μ μ
 μ μ μ Helicobacter pylori μ [462].
 , μ μ μ -
 [463]. μ μ
 G -384 μ μ μ
 μ μ (graft-versus-host-disease) μ
 μ [464]. , μ
 μ μ
 IL-2 μ μ
 [465].

6 (IL-6)

μ
 μ .
 IL-6. CNTF
 (ciliary neurotrophic factor), LIF (leukemia inhibitory
 factor), -1 (CT-1), GPA (growth promoting
 activity), IL-6 IL-11 [466]. IL-6,
 20-24KDa μ - μ μ . μ μ
 μ μ 30%.
 μ μ μ .

: μ IL-6

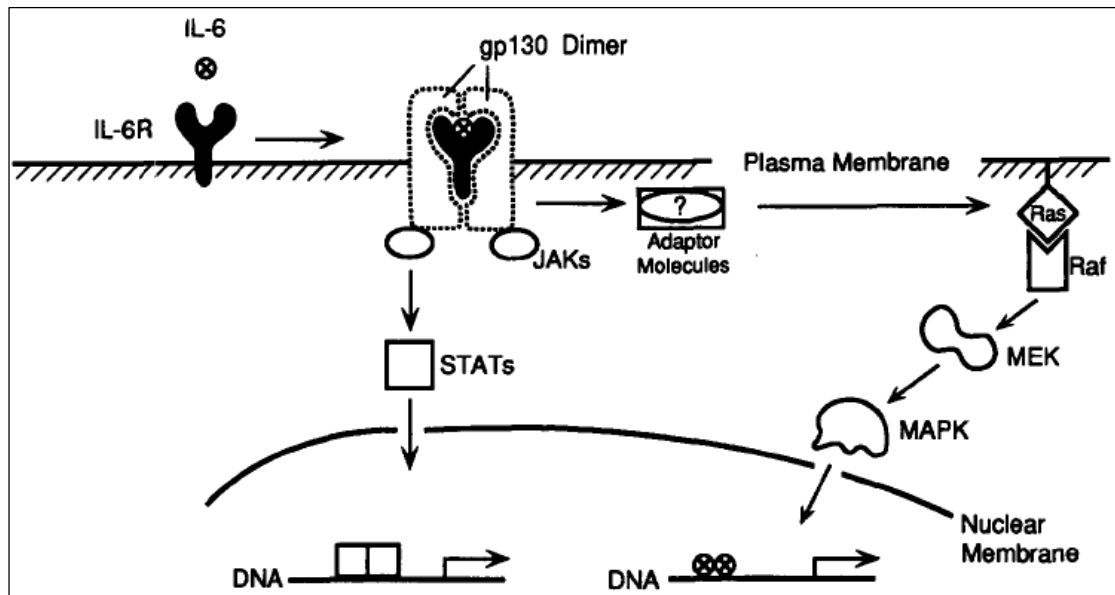


μ
 $\mu\mu$. μ μ -
 μ μ μ
 [467].

μ μ IL-6

IL-6
 μ
 μ μ - - -
 (WSXWS) μ μ .
 μ III (fibronectin III) [466].

IL-6 binds to the IL-6 receptor (IL-6R) and gp130. This complex recruits Janus Kinases (JAKs) and Signal Transducers and Activators of Transcription (STATs). The JAK-STAT pathway leads to transcription of target genes. Additionally, gp130 recruits adaptor molecules that activate Ras, which then triggers the Raf-MEK-MAPK cascade, leading to transcription of another set of target genes.



IL-6 binds to IL-6R and gp130. This complex recruits JAKs and STATs. The JAK-STAT pathway leads to transcription of target genes. Additionally, gp130 recruits adaptor molecules that activate Ras, which then triggers the Raf-MEK-MAPK cascade, leading to transcription of another set of target genes.

IL-6

μ μ μ μ IL-6
, μ μ
 μ [471-473]. μ ,
 μ μ IL-6 μ .
 μ , μ IL-6/
 μ μ gp130 μ - ,
 μ μ μ IL-6. μ ,
gp130 μ μ ,
IL-6, μ μ
IL-6/IL-6 (trans-signalling pathway).

IL-6

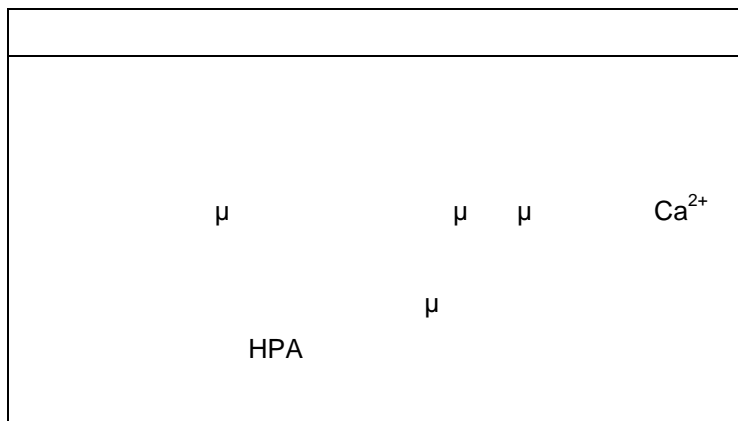
IL-6 μ
[90, 474-478]. μ mRNA
IL-6 μ , ,
[476]. , IL-6
[478]. mRNA IL-6
 μ
[474, 475]. μ , ,
 μ μ μ ,
 μ μ mRNA
mRNAs -
IL-6 μ μ ,

μ IL-6

_____ : μ μ mRNA IL-6
IL-1 TNF- . ,
- - μ IL-1RA
 μ TNF- , [479].
_____ :
IL-6 [480]. ,
 μ IL-6 . μ , IL-1 ,

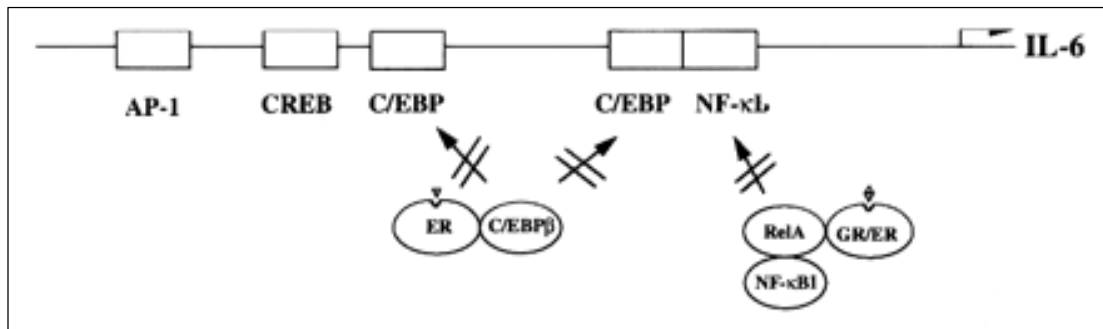
vivo μ IL-6 μ [495]. μ , in
 NMDA (N-methyl-D-aspartate)
 μ [131]. μ μ IL-6
 IL-6 μ μ μ Ca^{2+} μ
 [496, 497].
 _____: μ in vitro μ , IL-6
 μ NGF (nerve growth factor) [498].
 IL-6 μ NGF ,
 [499-501].
 _____: IL-6 μ
 in vitro [502] in vivo [133, 503-505]. , IL-6
 IL-6
 NGF [506].
 IL-6 μ μ _____: IL-6 μ
 [178].
 μ (μ IL-1 , TNF-) [179, 180]
 μ - - (HPA axis: hypothalamic-pituitary-adrenal axis), μ -
 μ , ,
 [507].
 IL-6 _____: IL-6
 μ , / IL-
 6 μ μ μ μ
 [508].

:
IL-6



IL-6

IL-6
 μ μ 7, p21-p15. 5 4 ,
 μ μ μ [509]. μ ,
 IL-6 μ NF-IL6
 (C/EBP), C/EBP , NF- B, μ
 μ [510, 511].



1: μ μ IL-6. -
 (glucocorticoid receptor) μ Re1A μ IL-6, μ GR -
 (estrogen receptor)/C/EBPbeta ER/NF- B μ IL-6, μ μ ER -
 . AP-1:AP-1 site, CREB:cyclic adenosine 5'-monophosphate-response element binding site, C/EBP:C/EBP binding site, NF- B:NF- B binding site.
 μ .

IL-6 (IL-6R)
 μ μ μ 1, q21-q22.
 μ μ IL-6
 μ μ (SNP)
 IL-6, -174 [512]. μ -
 μ IL-6 μ in vivo [369, 513]
 μ μ IL-6 [514]. 2 :
 C (), G ().
 CC μ μ IL-6 μ
 μ , GG IL-6

μ [515, 516]. , μ μ C
 IL-6 [517], μ
 IL-6 μ CC [518].
 μ . μ ,
 μ - μ , G μ μ
 0.87-1.0, μ μ 0.54-0.62 [519-521].
 , GG μ -
 μ (80-100%), μ
 μ (30-45%). , μ μ
 μ GG
 (38%), μ CC
 (18%) [512].

μ μ IL-6 μ

μ μ IL-6
 / μ .
 IL-6, μ .

<p>IL6-174G</p> <p>✓ μ 2 [522]</p> <p>✓ μ μ μ [523]</p> <p>✓ μ μ μ [524]</p> <p>✓ μ μ [525]</p> <p>✓ μ μ μ [295]</p> <p>✓ μ μ , μ μ [526]</p> <p>✓ μ μ - μ μ [527]</p> <p>✓ μ μ μ μ Kaposi μ μ μ HIV (Human Immunodeficiency Virus) [528]</p> <p>[529] μ μ Hodgkin</p>
<p>IL6-174C</p> <p>✓ μ μ [512]</p> <p>✓ μ μ / μ μ [530]</p> <p>✓ μ μ μ [531]</p>
<p>: μ μ IL-6 (-174G/C)</p>

μ , μ μ
 μ
 .
 μ μ μ
 μ μ IL-1 (-511C/T: **rs16944**), IL-1
 1 (+3953C/T: **rs1143634**), IL-1 (-889C/T: **rs1800587**), IL-1RA (VNTR 2
 : **rs2234663**), IL-2(+114G/T: **rs2069763**), IL-2(-384T/G: **rs2069762**)
 IL-6(-174G/C: **rs1800795**) (,
 μ) μ (μ
 , μ μ) .

μ 365 μ ,
 μ μ μ
 μ .

μ :
 1. μ ,
 , , (μ , μ ,
).
 >140mm Hg >90mm Hg μ μ
 μ μ .
 μ μ μ
 >126mg/dl μ
 μ μ >6.4%. μ
 μ μ >200mg/dl μ
 . μ μ
 :

2. μ μ
 μ μ μ
 (Glasgow Coma Scale: GCS). GCS μ μ 3 μ
 15 .
 μ μ μ

μ GCS: 3-8, 9-12 13-15, ,
μ

3. μ μ .

4. μ μ μ , μ μ μ -
μ , μ , μ μ ,
μ , μ μ
(, ,) μ
μ μ μ ABC/2.

5. μ μ
μ μ .

6. μ μ μ DN (μ
) μ μ

DNA μ μ .

7. μ 6 μ μ μ μ
(Glasgow Outcome Scale: GOS).
μ : (GOS= μ
(G) μ (MD)), -
μ μ μ
(GOS= μ (SD)
(V) (D)).

μ μ

μ DNA μ μ μ
μ [532]. -

DNA.

μ μ IL-1RA

μ PCR VNTR μ μ -
IL-1RA. μ μ [533]
28681 μ 28970 (Gene Bank Accession Number U65590).
μ μ (2%)
μ (EtBr) μ .

μ μ IL-1 (-511)

μ μ -511
IL-1 . PCR μ
(2.5%) μ (EtBr) μ
PCR μ
μ Ava I μ μ DNA
(2.5%) μ (EtBr)
UV.

μ μ IL-1 (+3953)

μ μ 5⁰
IL-1 . PCR μ
(1.5%) μ (EtBr) μ
PCR μ
μ Taq I μ μ DNA
(2%) μ (EtBr)
UV.

μ μ IL-1 (-889)

μ DNA μ μ
μ (PCR)
μ μ -889 IL-1 .
PCR μ (4%) μ
(EtBr) μ
PCR μ μ Nco I μ μ DNA

(4%)
 μ (EtBr) UV.

IL-2(-384)
 μ μ
 μ μ (PCR)
 μ μ -384 IL-2.
 PCR μ (3.5%)
 μ (EtBr) μ
 PCR μ Bfa I
 μ μ DNA
 (3.5%) μ (EtBr) UV.

IL-2(+114)
 μ μ
 IL-2 +114. μ μ PCR μ
 (2%) μ (EtBr)
 μ PCR
 μ μ Mwo I μ μ DNA
 (3%) μ
 (EtBr) UV.

IL-6(-174)
 μ μ DNA μ μ
 μ (PCR)
 μ μ -174 IL-6.
 PCR μ (2%) μ
 (EtBr) μ
 PCR μ SfaN I μ μ DNA
 (3%)
 μ (EtBr) UV.

Hardy-Weinberg (H-W) test online (power analysis) Fisher exact test (CaTS Power Calculator for Genetic Studies (Center for Statistical Genetics, University of Michigan, Ann Arbor, MI <http://ihg.gsf.de/cgi-bin/hw/hwa1.pl>).

(GCS 3-8, 9-12, 13-15, (logistic regression), (odds ratio) 95% (95% confidence intervals). GCS (3-8, 9-12, 13-15) IL-1 [IL-1 -889C/T (rs1800587), IL-1 +3953C/T (rs1143634), IL-1 -511C/T (rs16944), IL-1RA VNTR (rs2234663)] IL-2 [IL-2 +114G/T (rs2069763), IL-2 -384T/G (rs2069762)]

μ μ 4. μ μ 174G/C (rs1800795) IL-6
 μ μ 7, μ . μ μ μ
 μ (maximum-likelihood estimation). -
 μ μ shesis (<http://analysis.bio-x.cn/SHEsisMain.htm>),
 permutation tests , μ
 , μ Freeman-Tukey proportion tests. μ
 μ (D r²) μ
 μ μ μ μ μμ JLIN
 (<http://www.genepi.com.au/projects/jlin>).
 μ
 SPSS v.12.0 (Chicago, IL) SAS v.6.12 (Cary, NC). μ
 μ μ Bonferroni
<http://www.quantitativeskills.com/sisa/calculations/bonfer.htm>,

μ DNA
 μ DNA , μ ,
 μ μ DNA μ .
DNA μ DNA
 μ μ μ μ μ
 μ DNA.
 μ ,
 μ DNA μ
 μ μ μ μ μ
 μ DNA μ
 μ (μ). μ
10ml μ , Na₂EDTA.
 μ μ μ DNA
:

- μ
- Lysis I: NH₄Cl (155mM), KHCO₃ (10mM), EDTA (1mM), pH=7.4
- Lysis II: Tris (10mM), NaCl (400mM), di-Na EDTA (2mM), pH=8.2
- μ (10mg/ml)
- μ 10% w/v SDS (sodium dodecyl sulfate:)
- μ μ NaCl (6M)
-
- CH₃COONa (3M)
- 1%

μ DNA μ μ

:

1 μ

1. 12ml μ ddH₂O (μ). 15ml μ (μ).
2. μ 15min, 3500rpm, 4°C (μ).
3. , μ μ ddH₂O 12ml μ vortex.
4. μ 15min, 3500rpm, 4°C.
5. μ μ μ μ μ . μ μ lysis μ 12ml.
6. μ 15min, 3500rpm, 4°C.
7. μ μ 5 6 μ lysis .
8. μ μ μ μ μ . μ μ lysis I μ 6ml (μ lysis I lysis I μ μ DNA. , EDTA μ (Ca²⁺, Mg²⁺) , μ DNA). μ μ 15-30min, μ μ .
9. μ 750μl SDS (10% w/v) 100μl (10mg/ml) (SDS μ μ , μ μ μ μ . , μ μ DNA). μ μ 15-30min, μ μ .
10. 37°C 16-18hrs (overnight-O/N).

2 μ

1. μ 2ml 6M μ μ NaCl. μ 15sec (μ μ μ μ).
2. μ 15min, 3500rpm, 4°C.

3. μ DNA. μ μ
4. μ 15min, 3500rpm, 4°C.
5. μ μ 50ml μ
6. μ , μ 2 (2 μ μ)
 μ) CH₃COONa μ 1/10 μ
 μ . μ μ
 μ CH₃COONa
7. -20°C O/N.

3 μ

1. μ 50min, 3500rpm, 4°C.
2. μ μ . DNA « » μ μ
70% eppendorf (μ μ
 μ μ
 μ).
3. μ 30min, 12000rpm, 4°C.
4. μ μ 2 , μ
20min.
5. μ μ () , (μ μ μ -
DNA) μ μ μ , μ μ μ
 μ (μ μ)
6. μ .
7. DNA 4°C () -20°C (μ
 μ).

μ DNA μ DNA
 (1%) μ (EtBr) DNA μ
 UV.
 DNA.

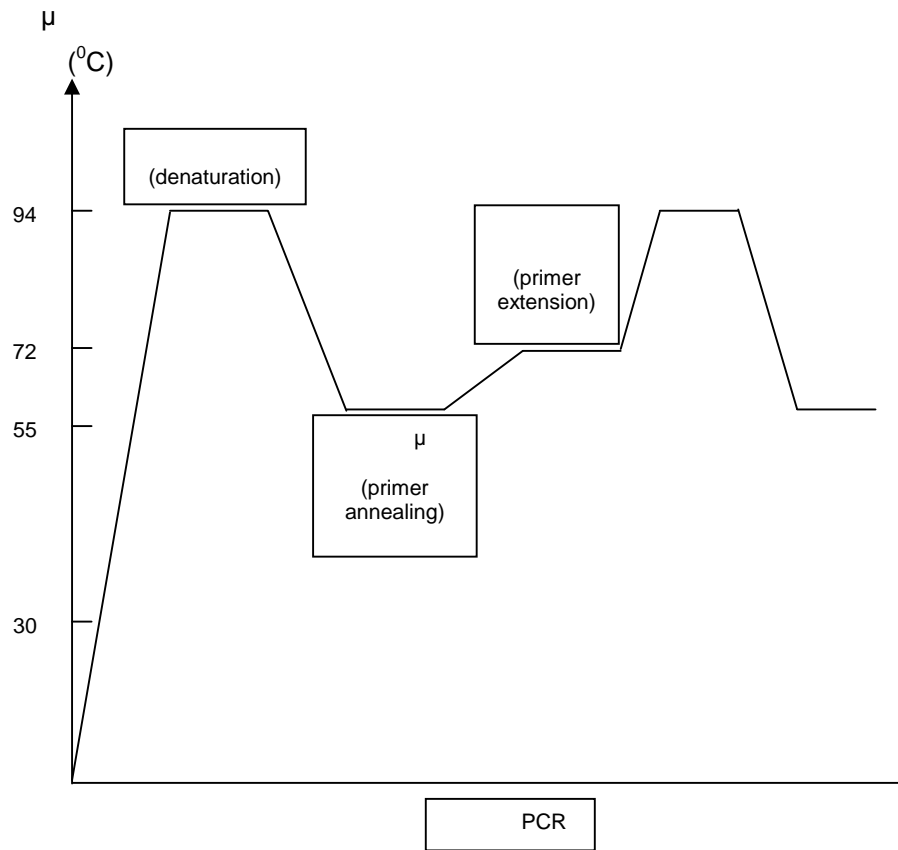
μ DNA μ DNA
 μ μ μ μ μ μ .
 DNA
 μ μ 260nm.
 μ , μ (OD) μ DNA
 (μ μ DNA 1:100) μ μ 260nm 280 nm
 μ μ 1cm. 260nm
 μ , 280 nm
 μ , μ .
 DNA μ μ OD₂₆₀/OD₂₈₀ μ
 μ 1,7 1,9 DNA, μ μ
 1,7 μ DNA , μ
 μ 1,9 μ μ RNA.
 μ μ 260nm μ OD μ 1
 50 μ g/ml DNA. μ μ
 DNA μ μ :

$\text{DNA } (\mu\text{g/ml}) = \quad \times 50 \times \mu \text{ OD}_{260}$
--

μ μ DNA
 μ 0,2 μ g DNA/ml μ .

μ (PCR: Polymerase Chain Reaction)

μ in vitro μ DNA (DNA-)
μ . PCR μ
DNA: DNA μ
μ μ DNA μ μ -
DNA
Taq DNA μ . μ DNA
μ μ μ DNA.
μ μ μ DNA
μ μ μ ,
μ μ μ
(primer). μ ,
μ μ DNA
- . μ μ
μ DNA μ μ -
PCR μ μ .
PCR μ μ
in vitro eppendorf μ μ
1 : μ DNA- μ , DNA
μ μ (denaturation)
2 : , μ
μ μ DNA- (primer annealing)
3 : DNA, Taq DNA μ dNTPs,
μ μ 3 μ
DNA- μ
(extension). DNA 5→3.
μ μ μ μ μ μ μ PCR
μ DNA, μ μ μ
μ μ μ μ DNA- μ
μ μ .



PCR (thermal cycler),
PCR. DNA.
G+C. DNA-
G+C 3
(2)

μ 94-95°C 3min
 DNA- μ
 μ 94 μ 95°C 30 μ 60
 sec. μ μ μ -
 DNA- / PCR μ μ
 μ μ DNA. , μ / μ -
 Taq DNA μ

 μ μ μ 5°C μ μ (T_m)
 65°C, μ μ μ 55 μ -
 μ μ μ μ -
 μ - μ DNA μ μ μ -
 3 μ μ μ -
 μ μ DNA. μ μ -

 μ μ μ μ μ
 72°C. μ Taq DNA μ
 35-100 μ μ μ pH, -
 μ μ μ μ DNA. T
 μ μ DNA- μ μ 1 μ 7 min,
 μ μ μ 1 min ($>1Kb$). ,
 μ μ μ μ ,
 μ .

μ :

➤	DNA	μ	(Taq DNA	μ)
➤			,	μ	-
	μ	-	(primers)		
➤		μ	5		-
			(dNTPs)		
➤			μ	MgCl ₂	
➤	μ		μ	Taq DNA	μ
➤		DNA		μ	- μ

μ , :

μ DNA- PCR

μ DNA-

μ DNA. , μ

DNA, μ PCR. -

μ ,

μ DNA

μ DNA- . ,

μ G+C, μ 40% μ 60%.

μ , μ 15 μ 30

μ μ 3 .

μ μ 0.1-0.2μ .

μ - μ μ (primer-

dimers) μ DNA- . ,

μ DNA- μ

μ (T_m, melting temperature), μ , μ

(T_a, annealing temperature). μ

DNA μ

μ μ

μ :

$m = (4^* \mu \text{ G+C}) + (2^* \mu \text{ A+T})$

μ T_m μ μ 55-65°C.
Taq DNA- μ μ μ ,
 Thermus aquaticus, μ μ μ
 DNA,
 μ μ PCR. , μ ,
 μ μ μ μ μ
 Taq DNA μ
 μ 1-5U/100 μ l. μ μ -
 μ , μ
 μ (temperature optimum) μ 72°C.
 , 5 (dNTPs) -
 DNA. dNTPs (dATP, dTTP, dCTP dGTP)
 μ μ μ
 μ , μ μ
 μ . ,
 dNTPs μ 20-200 μ .
 PCR μ (Mg^{2+}).
 Mg^{2+} μ - ,
 μ DNA μ .
 Mg^{2+} 1,5-2mM.
 Mg^{2+} μ
 μ .
 , μ μ ,
 PCR μ μ (buffer). μ μ
Tris-HCl pH μ 8,3 8,9.
 μ μ μ IL-1RA VNTR
 μ μ μ μ -
 :

Primer F: 5' **GCC CCT CAG CAA CAC TCC TAT** 3'

Primer R: 5' **CCC TGG TCC CCA CAA AAC TA** 3'

PCR

:

PCR _____ :	
95°C	7min
95°C	30sec
56°C	30sec
72°C	90sec
72°C	7min
4°C	∞

DNA (200ng/μl)	2.50 μl	0.02μgr
10× PCR buffer	2.50μl	1x
dNTPs (40mM)	0.50μl	0.8mM
Primer F (10μM)	0.25μl	0.1μ
Primer R (10μM)	0.25μl	0.1μ
MgCl ₂ (50mM)	1.00μl	2mM
Taq DNA Polymerase (5u/μl)	0.25μl	0.05u
dd ₂	17.75μl	-
(V .)	25.00μl	

50μl.



Primer F: 5' **TGG CAT TGA TCT GGT TCA TC** 3'
 Primer R: 5' **GTT TAG GAA TCT TCC CAC TT** 3'

PCR

:

PCR _____ :	
95°C	3min
95°C	1min
55°C	1min
74°C	1min
74°C	6min
4°C	∞

50μl.

DNA (200ng/μl)	5.00 μl	0.02μgr
10× PCR buffer	5.00μl	1x
dNTPs (40mM)	1.50μl	1.2mM
Primer F (10μM)	1.00μl	0.2μ
Primer R (10μM)	1.00μl	0.2μ
MgCl ₂ (50mM)	2.00μl	2mM
Taq DNA Polymerase (5u/μl)	0.50μl	0.05u
dd ₂	34.00μl	-
(V .)	50.00μl	

μ μ IL-1 +3953
 (+3953C/T) μ μ μ
 :

Primer F: ⁵ **GCC ACA GTG AGC AGT AAT AGA** ³
 Primer R: ⁵ **CAA CAA CAG GAA GAG AGA ACC** ³

PCR :

PCR :	
95°C	7min
95°C	30sec
55°C	30sec
72°C	90sec
72°C	7min
4°C	∞

} 32

50μl.

DNA (200ng/μl)	5.00 μl	0.02μgr
10× PCR buffer	5.00μl	1x
dNTPs (40mM)	1.50μl	1.2mM
Primer F (10μM)	1.00μl	0.2μ
Primer R (10μM)	1.00μl	0.2μ
MgCl ₂ (50mM)	2.00μl	2mM
Taq DNA Polymerase (5u/μl)	0.50μl	0.05u
dd ₂	34.00μl	-
(V .)	50.00μl	

μ μ IL-1 -889 (-889C/T)
 μ μ μ

:

Primer F: ⁵ **GTT CTA CCA CCT GAA CTA GGC**³

Primer R: ⁵ **TTA CAT ATG AGC CTT CCA TG**³

PCR

:

PCR _____:	
94°C	7min
94°C	1min
50°C	1min
72°C	1min
72°C	10min
4°C	∞

} 35

50μl.

DNA (200ng/μl)	5.00 μl	0.02μgr
10× PCR buffer	5.00μl	1x
dNTPs (40mM)	1.00μl	0.8mM
Primer F (10μM)	1.00μl	0.2μ
Primer R (10μM)	1.00μl	0.2μ
MgCl ₂ (50mM)	1.50μl	1.5mM
Taq DNA Polymerase (5u/μl)	0.50μl	0.05u
dd ₂	35.00μl	-
(V .)	50.00μl	

μ μ IL-2 -384 (-384T/G)
μ μ μ -

:

Primer F: ⁵CTC TTT GTT ACA TTA GCC CAC ACT TAG GTG³

Primer R: ⁵CAC AAT ATG CTA TTC ACA TGT TCA GTG TAG³

PCR

:

PCR		:
94 ⁰ C	2min	
94 ⁰ C	20sec	} 35
66 ⁰ C	40sec	
72 ⁰ C	20sec	
72 ⁰ C	10min	
4 ⁰ C	∞	

,

50μl.

DNA (200ng/μl)	5.00 μl	0.02μgr
10× PCR buffer	5.00μl	1x
dNTPs (40mM)	1.50μl	1.2mM
Primer F (10μM)	1.00μl	0.2μ
Primer R (10μM)	1.00μl	0.2μ
MgCl ₂ (50mM)	3.00μl	3mM
Taq DNA Polymerase (5u/μl)	0.50μl	0.05u
dd ₂	33.00μl	-
(V .)	50.00μl	

$\mu \mu$ IL-2 +114
 (+114G/T) $\mu \mu$ μ

Primer F: ⁵ **ATG TAC AGG ATG CAA CTC CT**³

Primer R: ⁵ **TGG TGA GTT TGG GAT TCT TG**³

PCR :

PCR :	
94 ⁰ C	2min
94 ⁰ C	20sec
52 ⁰ C	40sec
72 ⁰ C	20sec
72 ⁰ C	10min
4 ⁰ C	∞

50μl.

DNA (200ng/μl)	5.00 μl	0.02μgr
10× PCR buffer	5.00μl	1x
dNTPs (40mM)	1.50μl	1.2mM
Primer F (10μM)	1.00μl	0.2μ
Primer R (10μM)	1.00μl	0.2μ
MgCl ₂ (50mM)	3.00μl	3mM
Taq DNA Polymerase (5u/μl)	0.50μl	0.05u
dd ₂	33.00μl	-
(V)	50.00μl	

μ μ IL-6 -174 (-174G/C)
 μ μ μ

Primer F: 5' **CAG AAG AAC TCA GAT GAC TGG** 3'

Primer R: 5' **GCT GGG CTC CTG GAG GGG** 3'

PCR :

PCR _____ :	
95°C	5min
95°C	1min
63°C	1min
72°C	2min
72°C	10min
4°C	∞

} 30

25μl.

DNA (200ng/μl)	2.50 μl	0.02μgr
10× PCR buffer	2.50μl	1x
dNTPs (40mM)	0.50μl	0.8mM
Primer F (10μM)	0.50μl	0.2μ
Primer R (10μM)	0.50μl	0.2μ
MgCl ₂ (50mM)	0.75μl	1.5mM
Taq DNA Polymerase (5u/μl)	0.50μl	0.1u
dd ₂	17.25μl	-
(V)	25.00μl	

$v = E \times z / f$

DNA, μ

DNA, μ

O D- 3,6- -L- μ 1 \rightarrow 4 μ

μ

μ

Tris- -EDTA (1 \times : 10mM Tris, 0.09M
 , 0.5M EDTA) μ pH 8, μ

μ

μ

() μ

() μ

() μ DNA μ

(μ : 100mM EDTA , 20% w/v Ficoll, 1%Bromophenol blue).

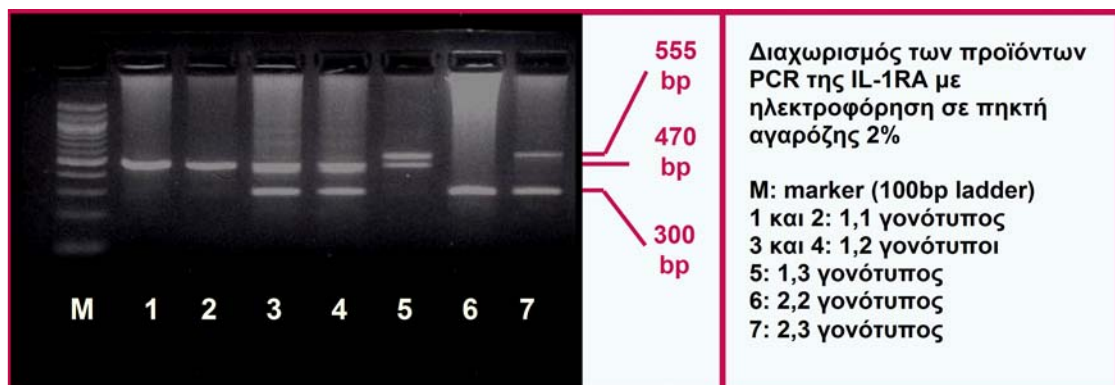
(« ») μ

(90-100V) μ DNA , DNA
 DNA μ
 pH, μ
 DNA
 DNA μ
 μ μ
 μ μ
 DNA.
 UV, DNA ,
 μ
 DNA.
 μ μ
 DNA μ μ , ,
 PCR

IL-1RA

T PCR μ (2%)
 μ (1μg/ml) μ
 μ PCR μ
 μ μ DNA (100bp DNA Ladder).
 μ μ :

Allele 1	4 repeats of 86bp	470bp
Allele 2	2 repeats	300bp
Allele 3	5 repeats	555bp
Allele 4	3 repeats	385bp
Allele 5	6 repeats	740bp



IL-1 (-511)

To PCR μ (2.5%)
 μ (1μg/ml) μ
 μ PCR μ
 μ μ DNA (100bp DNA Ladder).
 PCR μ μ , 304bp. ,
 , μ
 (2.5%) UV.
 μ μ :

Allele 1	PCR 2 μ μ	190 114bp
Allele 2	μ PCR	304bp

IL-1 (+3953)

To PCR μ (1.5%)
 μ (1μg/ml) μ
 μ PCR μ
 μ μ DNA (100bp DNA Ladder).
 PCR μ μ , 731bp. ,
 , μ
 (2%) UV.
 μ μ :

Allele C	PCR 2 μ μ	497 234bp
Allele T	μ PCR	731bp

IL-1 (-889)

To PCR μ (4%)
 μ (1μg/ml) μ
 μ PCR μ
 μ μ DNA (174 DNA/Hinf I Marker).

PCR μ μ , 99bp. ,
 μ
 (4%) μ UV.
 μ μ :

Allele T	μ PCR	99bp
Allele C	PCR 2 μ μ	83 16bp

IL-2(-384)

To PCR μ (3.5%)
 μ (1 μ g/ml) μ .
 μ PCR μ
 μ μ DNA (Low molecular weight DNA
 ladder). PCR μ μ , 169bp. ,
 μ
 (3.5%) μ UV.
 μ μ :

Allele 1	μ PCR	169bp
Allele 2	PCR 2 μ μ	137 32bp

IL-2(+114)

To PCR μ (2%)
 μ (1 μ g/ml) μ .
 μ PCR μ
 μ μ DNA (Low molecular weight DNA
 ladder). PCR μ μ , 262bp. ,
 μ
 (3%) μ UV.
 μ μ :

Allele G	μ PCR	262bp
Allele T	PCR 2 μ μ	151 111bp

IL-6(-174)

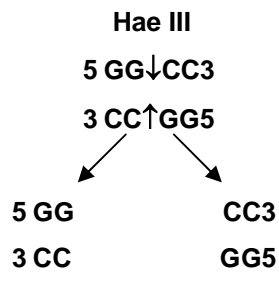
To μ PCR μ (2%)
 μ (1 μ g/ml) μ .
 μ PCR μ
 μ μ DNA (100bp DNA ladder).
PCR μ μ , 611bp. ,
, μ
(3%) UV.
 μ μ :

Allele G	μ PCR	611bp
Allele C	PCR 2 μ μ	367 244bp

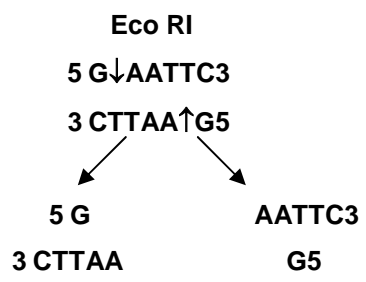
. μ μ
 μ DNA, μ μ 5 3 μ
 3 5. μ μ μ μ
 μ , μ Eco RI μ
 :



μ μ μ :
 > μ μ μ _____ (blunt or flush ends),
 μ : μ μ
 DNA. μ
 μ Hae III μ :



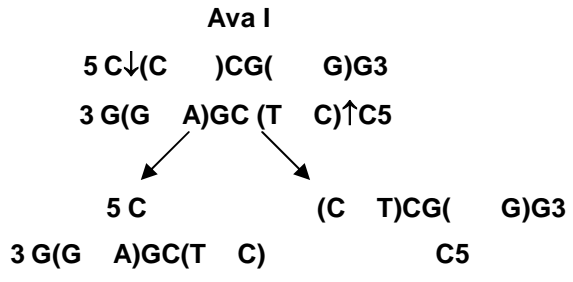
μ μ μ , μ μ μ μ 4 ,
 μ μ μ μ μ 5- - μ
 μμ 3- - μ μμ .
 > μ μ μ μ μ , μ μ
 : ' μ μ μ μ μ
 μ Eco RI μ μ μ :



μ μ , μ μ μ μ μ
 μ μ μ μ μ
 12 μ 15°C.

IL-1 (-511)

Ava I PCR μ μ μ Ava I. To
 variabilis. Ava I, μ μ Anabaena
 :

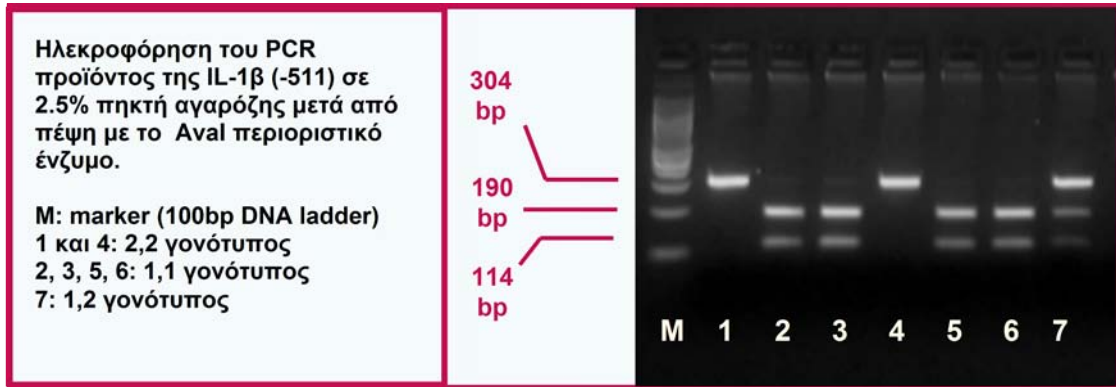


μ PCR
 40μl.

PCR product	30.00 μl	
μ Ava I (10u/μl)	1.00 μl	0.25u
10x NE Buffer 2	4.00 μl	1x
dd ₂	5.00 μl	-
(V)	40.00 μl	

16-18hrs (O/N) 37°C, μ μ DNA
 2.5%
 μ (1μg/ml) μ μ 100V
 UV. μ
 μ μ DNA (100bp)
 DNA Ladder).
 μ μ :

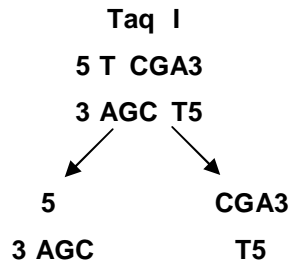
>IL1B-511*1 (-C -511
 IL-1B): μ Ava I μ
 PCR μ , 2 μ μ (190 114bp).
 >IL1B-511*2 (μ -T -511
 IL-1B): μ Ava I μ
 PCR (304bp).



IL-1 (+3953)

Taq I μ PCR μ Taq I.
 Escherichia Coli. μ
 μ - ,
 μ

Taq I, μ , μ
 :



μ PCR
 40μl.

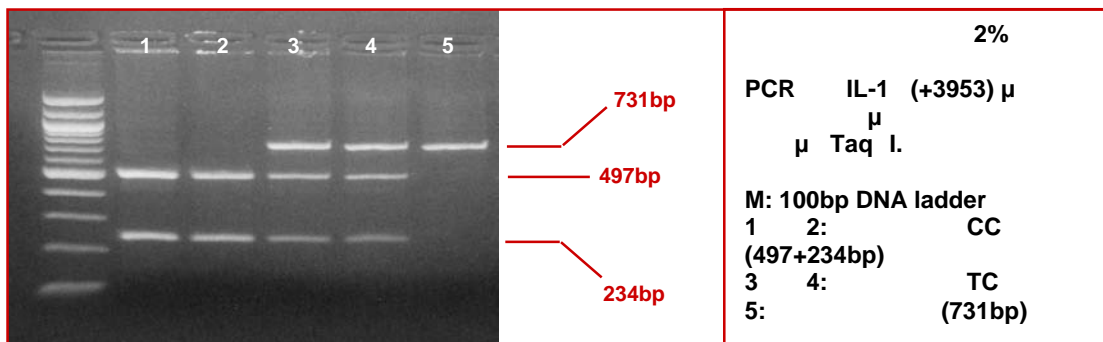
PCR product	30.00 µl	
µ Taq I (20u/µl)	1.00 µl	0.5u
10x NE Buffer 3	4.00 µl	1x
100x BSA*	0.40µl	1x
dd ₂	4.60 µl	-
(V)	40.00 µl	

* BSA (Bovine Serum Albumin: µ)
 µ , ,
 µ .
 16-18hrs (O/N) 65°C, µ µ DNA
 2%
 µ (1µg/ml) µ µ 100V
 UV. µ
 µ µ µ DNA (100bp
 DNA ladder).

µ µ :

➤ IL1B+3953*C (-C +3953 5
 IL-1B): µ Taq I µ
 PCR µ , 2 µ µ (497 234bp).

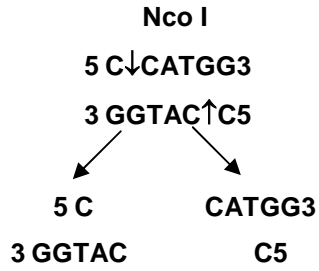
➤ IL1B+3953* (µ -T +3953 5
 IL-1B): µ Taq I µ
 PCR (731bp).



IL-1 (-889)

Nco I PCR µ µ µ Nco I. To
 µ µ Nocardia corallina.

Nco I, μ , μ
:



μ PCR
45 μ l.

PCR product	35.00 μ l	
μ Nco I (10u/ μ l)	1.50 μ l	0.33u
10x NE Buffer 4	4.50 μ l	1x
dd ₂	4.00 μ l	-
(V)	45.00 μ l	

16-18hrs (O/N) 37^oC, μ μ DNA

4%

μ (1 μ g/ml) μ μ 100V

UV. μ

μ μ μ DNA (174

DNA/Hinf I Marker).

μ μ :

μ IL1A-889*C (-C -889

IL-1A): μ Nco I μ

PCR μ , 2 μ μ (83 16bp).

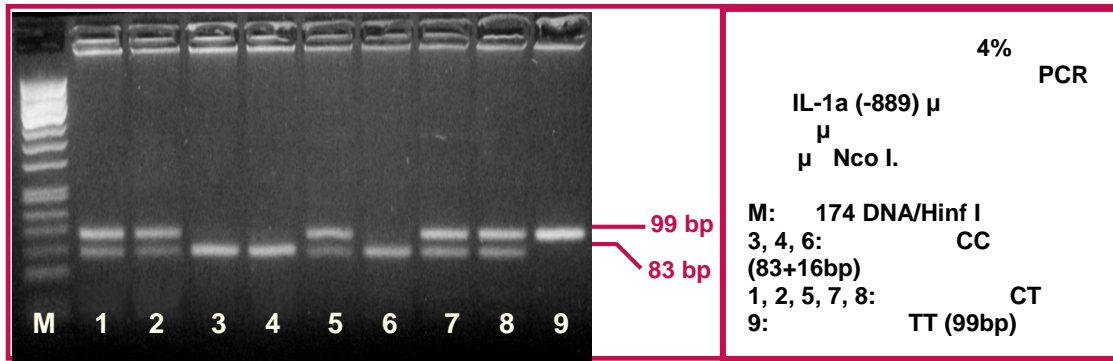
μ IL1A-889*T (-T -889

IL-1A): μ Nco I μ

PCR (99bp).

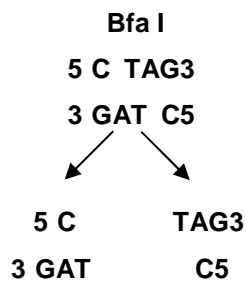
, μ μ 16bp , μ μ 83 bp

μ IL1A-889*C.



IL-2(-384)

PCR μ μ μ Bfa I. To Bfa
 I μ μ Bacteroides fragilis.
 Bfa I, μ , μ
 :

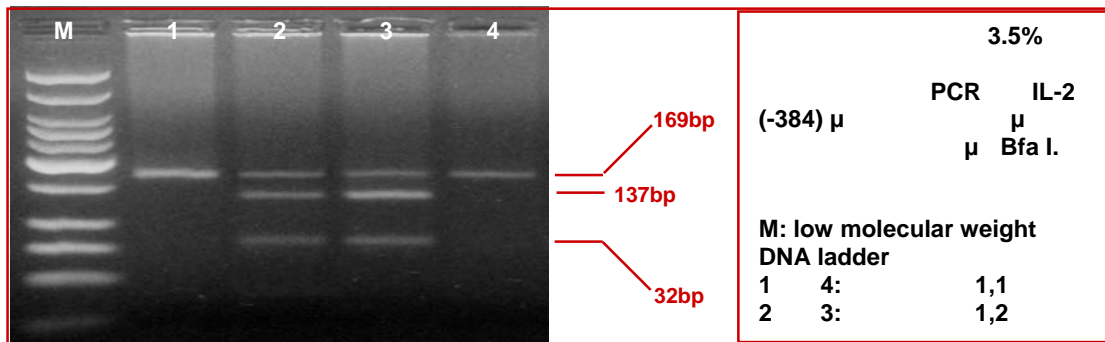


μ PCR
 40μl.

PCR product	30.00 μl	
μ Bfa I (5u/μl)	3.00 μl	0.38u
10x NE Buffer 4	4.00 μl	1x
dd ₂	3.00 μl	-
(V)	40.00 μl	

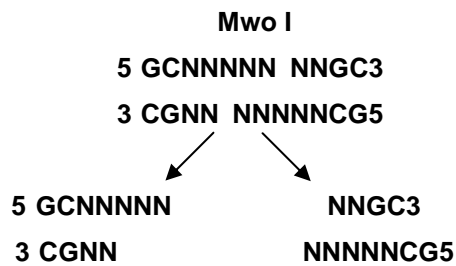
16-18hrs (O/N) 37°C, μ μ DNA
 3.5%
 μ (1μg/ml) μ μ 100V
 UV. μ μ
 μ μ DNA (Low
 molecular weight DNA ladder).

>IL2-384*1 (μ -T -384 μ IL-
 2): μ Bfa I μ PCR
 (169bp).
 >IL2-384*2 (-G -384 μ fa I μ PCR
 IL-2): μ μ μ (137 32bp).
 μ μ μ μ 32bp μ μ
 μ μ 137bp μ IL2-384*2.



IL-2(+114)

PCR μ μ μ Mwo I. To
 Mwo I μ μ Methanobacterium
 wolfeii. Mwo I, μ μ



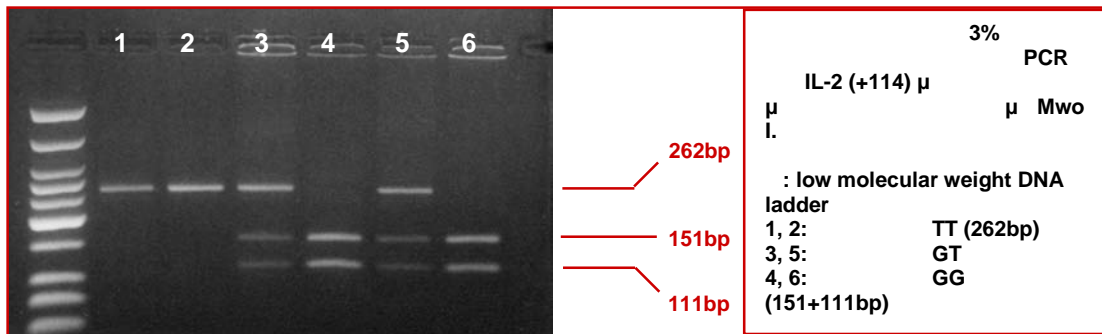
* :

μ PCR
 40μl.

PCR product	30.00 µl	
µ Mwo I (5u/µl)	1.00 µl	0.13u
10x NE Buffer 3	4.00 µl	1x
dd ₂	5.00 µl	-
(V)	40.00 µl	

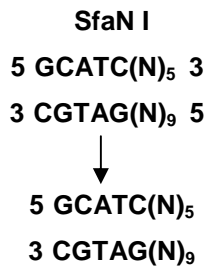
16-18hrs (O/N) 60°C, µ µ DNA
 3%
 µ (1µg/ml) µ µ 100V
 UV. µ µ
 µ µ µ DNA (Low
 molecular weight DNA ladder).

µ µ :
 >IL2+114*G (-G +114 1
 IL-2): µ Mwo I µ
 PCR µ , 2 µ µ (151 111bp).
 >IL2+114*T (µ -T +114 1
 IL-2): µ Mwo I µ
 PCR (262bp).



IL-6(-174)

SfaN I PCR µ µ µ SfaN I. To Streptococcus
 faecium. SfaN I, µ , µ



25µl.

µ

PCR

PCR product	20.00 µl	
µ SfaN I (1u/µl)	1.00 µl	0.04u
10x NE Buffer 3	2.50 µl	1x
dd ₂	1.50 µl	-
(V)	25.00 µl	

16-18hrs (O/N)

37°C,

µ µ

DNA

3%

µ

(1µg/ml) µ

µ

100V

UV.

µ

µ

µ

µ

DNA (Low

molecular weight DNA ladder).

µ

µ

:

➤ IL6-174*G (

-G

-174

IL-6):

µ SfaN I

µ

PCR (611bp).

➤ IL6-174*C (

-C

-174

IL-6):

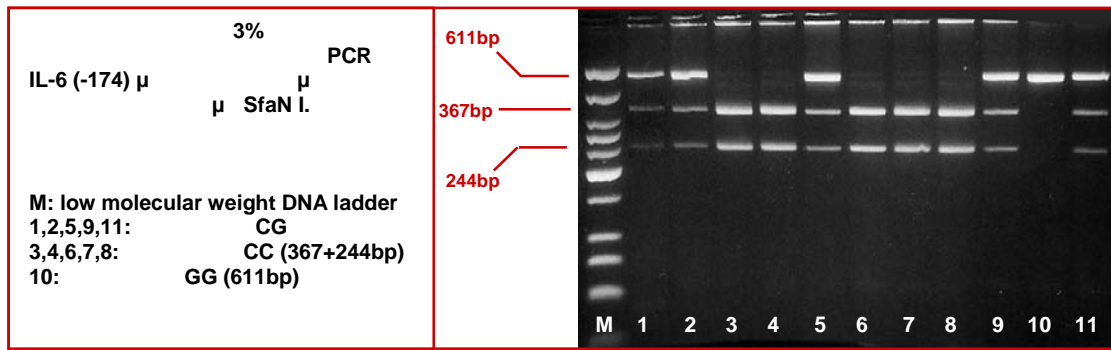
µ SfaN I µ

PCR

µ ,

2 µ µ

(367 244bp).



(mean) μ , 305 μ 60 (μ 5:1). μ 365
 μ 40.06 \pm 20,99 μ
 [1]. μ - μ
 μ μ μ (GCS). μ GCS
 μ 10.99 \pm 3.85. GCS
 34.24% (GCS 8), 15,34% μ
 μ 9-12 50.41%
 μ (GCS 13-15).
 μ μ μ .
 μ , 78 (21.37%) μ
 μ 287 (78.63%) μ .
 μ , 167 (45.75%) μ , 40 -
 (10.95%) μ , 185 (50.68%)
 μ , 91 (24.93%) μ μ ,
 24 (6.57%) μ 44
 (12.05%) μ .
 (GOS) μ 6 μ μ -
 μ , 242 (66.30%) , 54
 (14.79%) μ , 33 (9.04%)
 μ , 21 (5.75%)
 μ 15 μ (4.11%) μ 6 μ
 μ μ .
 μ , μ GOS 6 μ μ -
 μ μ . μ μ ,
 GOS, μ , μ 5 4 -
 μ 6 μ μ , μ
 3, 2 1 GOS. , , 296 μ

(81.09%) μ ,
 (18.91%) μ .

μ	μ	μ
	365	
	305 (83.6%)	
	60 (16.4)	
(±SD)	40.06±20.99	
(%)	57 (15,61%)	
(%)	17 (4.65%)	
	26 (7.12%)	
μ (%)	45 (12.32%)	
μ (%)	48 (13.15%)	
(%)	109 (29.86%)	
GCS (μ ±SD)	10.99±3.85	
GCS: 3-8	125 (34.24%)	
GCS: 9-12	56 (15.34%)	
GCS: 13-15	184 (50.41%)	
6μ		
(5)	242 (66.30%)	
(4)	54 (14.79%)	
(3)	33 (9.04%)	
(2)	21 (5.75%)	
(1)	15 (4.11%)	
(4-5)	296 (81.09%)	
μ (1-3)	69 (18.91%)	
μ		
μ	78 (21.37%)	
μ	287 (78.63%)	
μ	167 (45.75%)	
μ μ	40 (10.95%)	
μ	185 (50.68%)	
μ μ	55 (15.06%)	
μ μ	91 (24.9%)	

μ	24 (6.57%)
μ	44 (12.05%)
SD=	, GCS= μ μ

μ μ μ μ : IL-1 -511C/T (rs16944), IL-1 +3953C/T (rs1143634), IL-1 -889C/T (rs1800587), IL-1RA VNTR 2 (rs2234663), IL-2 +114G/T (rs2069763), IL-2 -384T/G (rs2069762) IL-6 -174G/C (rs1800795).

μ μ μ μ , μ μ μ μ Hapmap (<http://hapmap.ncbi.nlm.nih.gov>, Protocol: Phase3 _Draft2).

	SNP	μ rs	μ μ			Minor allele frequency μ
1.	IL-1 (-889C/T)	rs1800587	113259431 (chr 2)	IL-1	μ	0.252
2.	IL-1 (+3953C/T)	rs1143634	113306861 (chr 2)	5	Coding-synonymous	0.208
3.	IL-1 (-511C/T)	rs16944	113311338 (chr 2)	IL-1	μ	0.358
4.	IL-1RA (VNTR)	rs2234663	113604577 (chr 2)	2	No-coding	No data
5.	IL-2(+114G/T)	rs2069763	123596932 (chr 4)	1	Coding-synonymous	0.342
6.	IL-2(-384T/G)	rs2069762	123597430 (chr 4)	IL-2	μ	0.273
7.	IL-6(-174G/C)	rs1800795	22733170 (chr 7)	IL-6	μ	0.465

(1-7) μ - μ μ : μ , μ μ μ . $\mu\mu$ Fisher exact test μ μ Hardy-Weinberg (H-W) . μ μ IL-1 (-511C/T, rs16944), IL-1 (+3953C/T, rs1143634), IL-1 (-889C/T, rs1800587), IL-1RA (VNTR, rs2234663), IL-2(+114G/T, rs2069763), IL-6(-174G/C, rs1800795) H-W (p>0.05), μ μ μ μ

H-W (p<0.01).
 TG (59.18%), GG,
 GG [534].
 8-14
 GCS, 2)
 6μ GOS 3)
 (allele contrast).
 (logistic regression analysis)
 6μ GCS
 μ 7 SNPs μ Bonferroni
 0.05 0.007.
 μ ρ μ 0.007.
 IL-1 (-511C/T, rs16944), IL-1 (+3953C/T, rs1143634), IL-1 (-889C/T, rs1800587), IL-2 (-384T/G rs2069762) IL-6(-174G/C, rs1800795)
 IL-2(+114G/T, rs2069763)
 2 ()

. μ , μ μ μ μ μ μ
 μ GCS 3-8 μ GCS 13-15 μ μ μ
 ([μ μ] 0.57 [0.38-0.80], $p=0.002$)
 μ μ μ (0.53 [0.31-0.81], $p=0.009$). μ
 μ μ μ Bonferroni μ
 μ $p=0.02$ 2.32 μ μ
 μ 1.18 5.12.
 μ μ IL-1RA (VNTR, rs2234663)
 μ 2 (μ) 4.5 μ
 μ μ μ μ (μ μ)
 [μ μ] 4.57 [1.67-12.96], $p=0.004$). μ μ
 μ μ (0.26 [0.14-
 0.71], $p=0.003$).
 , μ 2 μ
 : GCS 3-8 μ GCS 9-12
 (μ 0.32 [0.12-0.90], $p=0.030$ μ
 0.32 [0.16-0.83], $p=0.02$) GCS 3-8 μ GCS 13-15
 (μ 0.37 [0.19-0.78], $p=0.02$ μ -
 μ 0.45 [0.25-0.91], $p=0.017$).
 , 6 μ , μ 2 μ
 : μ 0.39 [0.19-0.89], $p=0.011$
 μ μ 0.43 [0.19-0.86], $p=0.035$.

1: μ μ μ μ IL-1 (+3953C/T rs1143634)																
							μ									
							1 (C)	%	2 (T)	%						
							1.1 (CC)	%	1.2 (CT)	%	2.2 (TT)	%				
(n=365)							211	57,81%	125	34,25%	29	7,95%	547	74,93%	183	25,07%
(n=296)							172	58,11%	100	33,78%	24	8,11%	444	75,00%	148	25,00%
(n=69)							39	56,52%	25	36,23%	5	7,25%	103	74,64%	35	25,36%
GCS 3-8 (n=125)							76	60,80%	40	32,00%	9	7,20%	192	76,80%	58	23,20%
GCS 9-12 (n=56)							29	51,79%	21	37,50%	6	10,71%	79	70,54%	33	29,46%
GCS 13-15 (n=184)							106	57,61%	64	34,78%	14	7,61%	276	75,00%	92	25,00%
μ μ (n=287)							167	58,19%	96	33,45%	24	8,36%	430	74,91%	144	25,09%
μ (n=78)							44	56,41%	29	37,18%	5	6,41%	117	75,00%	39	25,00%

2: μ μ μ μ IL-2 (-384T/G rs2069762)											
							μ				
		1.1 (TT)	%	1.2 (TG)	%	2.2 (GG)	%	1 (T)	%	2 (G)	%
(n=365)		149	40,82%	216	59,18%	0	0,00%	514	70,41%	216	29,59%
(n=296)		120	40,54%	176	59,46%	0	0,00%	416	70,27%	176	29,73%
(n=69)		29	42,03%	40	57,97%	0	0,00%	98	71,01%	40	28,99%
GCS 3-8 (n=125)		56	44,80%	69	55,20%	0	0,00%	181	72,40%	69	27,60%
GCS 9-12 (n=56)		22	39,29%	34	60,71%	0	0,00%	78	69,64%	34	30,36%
GCS 13-15 (n=184)		71	38,59%	113	61,41%	0	0,00%	255	69,29%	113	30,71%
μ	μ (n=287)	116	40,42%	171	59,58%	0	0,00%	403	70,21%	171	29,79%
	μ (n=78)	33	42,31%	45	57,69%	0	0,00%	111	71,15%	45	28,85%

3: μ μ μ μ IL-2 (+114G/T rs2069763)										
							μ			
	1.1 (GG)	%	1.2 (GT)	%	2.2 (TT)	%	1 (G)	%	2 (T)	%
(n=365)	183	50,14%	145	39,73%	37	10,14%	511	70,00%	219	30,00%
(n=296)	150	50,68%	115	38,85%	31	10,47%	415	70,10%	177	29,90%
(n=69)	33	47,83%	30	43,48%	6	8,70%	96	69,57%	42	30,43%
GCS 3-8 (n=125)	51	40,80%	55	44,00%	19	15,20%	157	62,80%	93	37,20%
GCS 9-12 (n=56)	27	48,21%	23	41,07%	6	10,71%	77	68,75%	35	31,25%
GCS 13-15 (n=184)	105	57,07%	67	36,41%	12	6,52%	277	75,27%	91	24,73%
μ μ (n=287)	145	50,52%	112	39,02%	30	10,45%	402	70,03%	172	29,97%
μ (n=78)	38	48,72%	33	42,31%	7	8,97%	109	69,87%	47	30,13%

4: μ μ μ μ IL-6 (-174C/G rs1800795)											
							μ				
		1.1 (CC)	%	1.2 (CG)	%	2.2 (GG)	%	1 (C)	%	2 (G)	%
(n=365)		225	61,64%	126	34,52%	14	3,84%	576	78,90%	154	21,10%
(n=296)		181	61,15%	103	34,80%	12	4,05%	465	78,55%	127	21,45%
(n=69)		44	63,77%	23	33,33%	2	2,90%	111	80,43%	27	19,57%
GCS 3-8 (n=125)		81	64,80%	41	32,80%	3	2,40%	203	81,20%	47	18,80%
GCS 9-12 (n=56)		29	51,79%	26	46,43%	1	1,79%	84	75,00%	28	25,00%
GCS 13-15 (n=184)		115	62,50%	59	32,07%	10	5,43%	289	78,53%	79	21,47%
μ μ	(n=287)	177	61,67%	97	33,80%	13	4,53%	451	78,57%	123	21,43%
μ	(n=78)	48	61,54%	29	37,18%	1	1,28%	125	80,13%	31	19,87%

5: μ μ μ μ IL-1RN (VNTR rs2234663)																		
												μ						
μ	μ	IL-1RA	1.1	%	1.3	%	1.2	%	2.2	%	2.3	%	1	%	2	%	3	%
(n=151)			81	53,64%	6	3,97%	51	33,77%	12	7,95%	1	0,66%	219	72,52%	76	25,17%	7	2,32%
(n=119)			60	50,42%	3	2,52%	44	36,97%	11	9,24%	1	0,84%	167	70,17%	67	28,15%	4	1,68%
(n=32)			21	65,63%	3	9,38%	7	21,88%	1	3,13%	0	0,00%	52	81,25%	9	14,06%	3	4,69%
GCS 3-8 (n=59)			24	40,68%	1	1,69%	27	45,76%	7	11,86%	0	0,00%	76	64,41%	41	34,75%	1	0,85%
GCS 9-12 (n=24)			15	62,50%	2	8,33%	7	29,17%	0	0,00%	0	0,00%	39	81,25%	7	14,58%	2	4,17%
GCS 13-15 (n=68)			42	61,76%	3	4,41%	17	25,00%	5	7,35%	1	1,47%	104	76,47%	28	20,59%	4	2,94%
(n=117)	μ	μ	55	47,01%	4	3,42%	46	39,32%	11	9,40%	1	0,85%	160	68,38%	69	29,49%	5	2,14%
(n=34)		μ	26	76,47%	2	5,88%	5	14,71%	1	2,94%	0	0,00%	59	86,76%	7	10,29%	2	2,94%

6: μ μ μ μ IL-1 (-511C/T rs16944)										
							μ			
	1.1 (CC)	%	1.2 (CT)	%	2.2 (TT)	%	1 (C)	%	2 (T)	%
(n=151)	51	33,77%	83	54,97%	17	11,26%	185	61,26%	117	38,74%
(n=119)	42	33,33%	69	54,76%	15	11,90%	153	60,71%	99	39,29%
(n=32)	9	36,00%	14	56,00%	2	8,00%	32	64,00%	18	36,00%
GCS 3-8 (n=59)	20	33,90%	36	61,02%	3	5,08%	76	64,41%	42	35,59%
GCS 9-12 (n=24)	9	37,50%	11	45,83%	4	16,67%	29	60,42%	19	39,58%
GCS 13-15 (n=68)	22	32,35%	36	52,94%	10	14,71%	80	58,82%	56	41,18%
A μ (n=117)	36	30,77%	67	57,26%	14	11,97%	139	59,40%	95	40,60%
X μ (n=34)	15	44,12%	16	47,06%	3	8,82%	46	67,65%	22	32,35%

7: μ		μ		μ		μ		IL-1 (-889 C/T rs1800587)			
								μ			
		1.1 (CC)	%	1.2 (CT)	%	2.2 (TT)	%	1 (C)	%	2 (T)	%
(n=215)		107	49,77%	84	39,07%	24	11,16%	298	69,30%	132	30,70%
(n=185)		93	50,27%	72	38,92%	20	10,81%	258	69,73%	112	30,27%
(n=30)		14	46,67%	12	40,00%	4	13,33%	40	66,67%	20	33,33%
GCS 3-8 (n=64)		34	53,13%	24	37,50%	6	9,38%	92	71,88%	36	28,13%
GCS 9-12 (n=27)		9	33,33%	12	44,44%	6	22,22%	30	55,56%	24	44,44%
GCS 13-15 (n=124)		64	51,61%	48	38,71%	12	9,68%	176	70,97%	72	29,03%
μ μ (n=168)		82	48,81%	69	41,07%	17	10,12%	233	69,35%	103	30,65%
μ (n=47)		25	53,19%	15	31,91%	7	14,89%	65	69,15%	29	30,85%

8: IL-1 (+3953C/T rs1143634)				
[2] vs [1]				
	2 ()	1 (C)	OR (95%CI)	p-value
(n=296)	148	444	-	
(n=69)	35	103	1.03 [0.68-1.71] [†]	p=0.91
GCS 3-8 (n=125)	58	192	-	
GCS 9-12 (n=56)	33	79	1.35 [0.87-2.35]*	p=0.22
GCS 13-15 (n=184)	92	276	1.15 [0.69-1.73]*	p=0.65
μ (n=287)	144	430	-	
μ (n=78)	39	117	0.98 [0.68-1.53] [‡]	p=0.96
μ 2 [12+22] vs [11]				
	[12+22] (/-)	[11] (CC)	OR (95%CI)	p-value
(n=296)	124	172	-	
(n=69)	30	39	1.01 [0.68-1.84] [†]	p=0.82
GCS 3-8 (n=125)	49	76	-	
GCS 9-12 (n=56)	27	29	1.49 [0.73-2.77]*	p=0.28
GCS 13-15 (n=184)	78	106	1.16 [0.76-1.84]*	p=0.61
μ (n=287)	120	167	-	
μ (n=78)	34	44	1.03 [0.61-1.81] [‡]	p=0.75
μ 2 [22] vs [11+12]				
	[22] ()	[11+12] (C/-)	OR (95%CI)	p-value
(n=296)	24	272	-	
(n=69)	5	64	1.16 [0.43-3.03] [†]	p=0.84
GCS 3-8 (n=125)	9	116	-	
GCS 9-12 (n=56)	6	50	0.68 [0.27-1.98]*	p=0.46
GCS 13-15 (n=184)	14	170	0.99 [0.41-2.63]*	p=0.85
μ (n=287)	24	263	-	
μ (n=78)	5	73	1.37 [0.46-3.66] [‡]	p=0.54

(*)
GCS

μ . (†) , μ μ μ . (‡) μ GCS / μ

9: IL-2 (-384T/G rs2069762)				
[2] vs [1]				
	2 (G)	1 (T)	OR (95%CI)	p-value
(n=296)	176	416	-	
(n=69)	40	98	0.92 [0.68-1.90] [†]	p=0.84
GCS 3-8 (n=125)	69	181	-	
GCS 9-12 (n=56)	34	78	1.12 [0.79-1.81]*	p=0.52
GCS 13-15 (n=184)	113	255	1.15 [0.79-1.68]*	p=0.45
μ (n=287)	171	403	-	
μ (n=78)	45	111	0.98 [0.61-1.44] [†]	p=0.85
μ 2 [12+22] vs [11]				
	[12+22] (G/-)	[11] TT)	OR (95%CI)	p-value
(n=296)	176	120	-	
(n=69)	40	29	0.92 [0.59-1.83] [†]	p=0.87
GCS 3-8 (n=125)	69	56	-	
GCS 9-12 (n=56)	34	22	1.21 [0.63-2.37]*	p=0.46
GCS 13-15 (n=184)	113	71	1.45 [0.73-2.28]*	p=0.21
μ (n=287)	171	116	-	
μ (n=78)	45	33	0.91 [0.54-1.58] [†]	p=0.78
μ 2 [22] vs [11+12]				
	[22] (GG)	[11+12] (T/-)	OR (95%CI)	p-value
(n=296)	0	296	-	
(n=69)	0	69	0.21 [0.004-12.10] [†]	p=1.00
GCS 3-8 (n=125)	0	125	-	
GCS 9-12 (n=56)	0	56	0.42 [0.007-23.41]*	p=1.00
GCS 13-15 (n=184)	0	184	1.46 [0.035-77.63]*	p=1.00
μ (n=287)	0	287	-	
μ (n=78)	0	78	0.24 [0.004-16.45] [†]	p=1.00

(*)
GCS

μ . (†) μ μ μ . (†) μ GCS / μ

10: IL-2 (+114G/T rs2069763)				
[2] vs [1]				
	2 (T)	1 (G)	OR (95%CI)	p-value
(n=296)	177	415	-	
(n=69)	42	96	1.03 [0.67-1.57] [†]	p=0.93
GCS 3-8 (n=125)	93	157	-	
GCS 9-12 (n=56)	35	77	0.767 [0.477-1.234]*	p=0.27371
GCS 13-15 (n=184)	91	277	0.57 [0.38-0.80]*	p=0.002
μ (n=287)	172	402	-	
μ (n=78)	47	109	1.04 [0.67-1.51] [‡]	p=0.94
μ 2 [12+22] vs [11]				
	[12+22] (T/-)	[11] (GG)	OR (95%CI)	p-value
(n=296)	146	150	-	
(n=69)	36	33	1.17 [0.61-1.92] [†]	p=0.64
GCS 3-8 (n=125)	74	51	-	
GCS 9-12 (n=56)	29	27	0.72 [0.36-1.41]*	p=0.37
GCS 13-15 (n=184)	79	105	0.53 [0.31-0.81]*	p=0.009
μ (n=287)	142	145	-	
μ (n=78)	40	38	1.12 [0.68-1.79] [‡]	p=0.75
μ 2 [22] vs [11+12]				
	[22] (TT)	[11+12] (G/-)	OR (95%CI)	p-value
(n=296)	31	265	-	
(n=69)	6	63	1.27 [0.46-2.88] [†]	p=0.61
GCS 3-8 (n=125)	19	106	-	
GCS 9-12 (n=56)	6	50	1.45 [0.58-3.52]*	p=0.48
GCS 13-15 (n=184)	12	172	2.32 [1.18-5.12]*	p=0.02
μ (n=287)	30	257	-	
μ (n=78)	7	71	1.17 [0.48-2.55] [‡]	p=0.65

(*)
GCS

μ . (†) , μ μ μ . (‡) / μ

11: IL-6 (-174C/G rs1800795)				
[2] vs [1]				
	2 (G)	1 (C)	OR (95%CI)	p-value
(n=296)	127	465	-	
(n=69)	27	111	0.87 [0.52-1.55] [†]	p=0.68
GCS 3-8 (n=125)	47	203	-	
GCS 9-12 (n=56)	28	84	1.41 [0.87-2.32]*	p=0.19
GCS 13-15 (n=184)	79	289	1.167 [0.75-1.73]*	p=0.43
μ (n=287)	123	451	-	
μ (n=78)	31	125	0.93 [0.54-1.42] [‡]	p=0.68
μ 2 [12+22] vs [11]				
	[12+22] (G/-)	[11] (CC)	OR (95%CI)	p-value
(n=296)	115	181	-	
(n=69)	25	44	0.87 [0.53-1.59] [†]	p=0.66
GCS 3-8 (n=125)	44	81	-	
GCS 9-12 (n=56)	27	29	1.74 [0.92-3.01]*	p=0.096
GCS 13-15 (n=184)	69	115	1.12 [0.67-1.81]*	p=0.67
μ (n=287)	110	177	-	
μ (n=78)	30	48	1.05 [0.64-1.71] [‡]	p=0.96
μ 2 [22] vs [11+12]				
	[22] (GG)	[11+12] (C/-)	OR (95%CI)	p-value
(n=296)	12	284	-	
(n=69)	2	67	1.42 [0.32-5.88] [†]	p=0.63
GCS 3-8 (n=125)	3	122	-	
GCS 9-12 (n=56)	1	55	1.31 [0.12-12.96]*	p=0.76
GCS 13-15 (n=184)	10	174	0.43 [0.12-1.60]*	p=0.21
μ (n=287)	13	274	-	
μ (n=78)	1	77	3.63 [0.47-25.28] [‡]	p=0.19

(*)
GCS

μ . (†) , μ μ μ . (‡) / μ

12: IL-1RN (VNTR rs2234663)				
[2] vs [1]				
	2	1	OR (95%CI)	p-value
(n=119)	67	167	-	
(n=32)	9	52	0.43 [0.19-0.86][†]	p=0.035
GCS 3-8 (n=59)	41	76	-	
GCS 9-12 (n=24)	7	39	0.32 [0.16-0.83]*	p=0.02
GCS 13-15 (n=68)	28	104	0.45 [0.25-0.91]*	p=0.017
A μ (n=117)	69	160	-	
X μ (n=34)	7	59	0.26 [0.14-0.71][‡]	p=0.003
μ 2 [12+22] vs [11]				
	Allele 2 carrier	Allele 2 non-carrier	OR (95%CI)	p-value
(n=119)	56	63	-	
(n=32)	8	24	0.37 [0.19-0.78][†]	p=0.02
GCS 3-8 (n=59)	34	25	-	
GCS 9-12 (n=24)	7	17	0.32 [0.12-0.90]*	p=0.030
GCS 13-15 (n=68)	23	45	0.39 [0.19-0.89]*	p=0.011
A μ (n=117)	58	59	-	
X μ (n=34)	6	28	4.57 [1.67-12.96][‡]	p=0.004
μ 2 [22] vs [11+12]				
	[22]	[11+12+1.3+2.3]	OR (95%CI)	p-value
(n=119)	11	108	-	
(n=32)	1	31	2.83 [0.28-25.13] [†]	p=0.31
GCS 3-8 (n=59)	7	52	-	
GCS 9-12 (n=24)	0	24	6.31 [0.37-110.1]*	p=0.11
GCS 13-15 (n=68)	5	63	1.68 [0.514-5.52]*	p=0.47
A μ (n=117)	11	106	-	
X μ (n=34)	1	33	3.21 [0.53-25.6] [‡]	p=0.25

(*)
GCS

, μ .(†) μ μ μ .(†) μ GCS / μ

13: IL-1 (-511C/T rs16944)				
[2] vs [1]				
	2 (T)	1 (C)	OR (95%CI)	p-value
(n=119)	99	153	-	
(n=32)	18	32	0.82 [0.49-1.60] [†]	p=0.65
GCS 3-8 (n=59)	42	76	-	
GCS 9-12 (n=24)	19	29	1.17 [0.56-2.12]*	p=0.65
GCS 13-15 (n=68)	56	80	1.23 [0.77-1.98]*	p=0.38
A μ (n=117)	95	139	-	
X μ (n=34)	22	46	0.73 [0.36-1.11] [‡]	p=0.29
μ 2 [12+22] vs [11]				
	[12+22] (T/-)	[11] (CC)	OR (95%CI)	p-value
(n=119)	84	42	-	
(n=32)	16	9	0.87 [0.33-1.96] [†]	p=0.63
GCS 3-8 (n=59)	39	20	-	
GCS 9-12 (n=24)	15	9	0.86 [0.32-2.29]*	p=0.755
GCS 13-15 (n=68)	46	22	1.00 [0.48-2.10]*	p=0.993
A μ (n=117)	80	37	-	
X μ (n=34)	19	15	1.71 [0.78-3.73] [‡]	p=0.087
μ 2 [22] vs [11+12]				
	[22] (TT)	[11+12] (C/-)	OR (95%CI)	p-value
(n=119)	15	111	-	
(n=32)	2	23	1.51 [0.37-7.6] [†]	p=0.54
GCS 3-8 (n=59)	3	56	-	
GCS 9-12 (n=24)	4	20	0.23 [0.17-1.44]*	p=0.12
GCS 13-15 (n=68)	10	58	0.311 [0.151-1.22]*	p=0.10
A μ (n=117)	14	103	-	
X μ (n=34)	3	31	1.48 [0.31-4.88] [‡]	p=0.65

(*)
GCS

μ . (†) , μ μ μ . (†) μ GCS / μ

15 16. $\mu > 0.05$. μ μ μ
 SNPs) Bonferroni.
 IL-1 (IL-1 -889C/T, IL-1 +3953C/T, IL-1 -511C/T, IL-1RA VNTR)
 μ 6μ 6 μ μ
 $\mu > 0.05$. VI (2-1-1-1,
 μ SNP IL-1 -889 C/T μ)
 μ μ μ 6μ μ (
 [μ μ] 3.01 [1.59-5.69], $p=0.0004$). Global test
 μ ($p=0.0025$).
 μ μ GCS 3-8 μ GCS 9-12
 μ μ . IV (1-1-2-2) μ μ -
 (0.16 (0.05-0.51), $p=0.0005$),
 μ VIII (2-2-2-1) μ μ
 GCS 9-12 (9.79 (2.74-34.98), $p=2.64 \cdot 10^{-5}$). Global test
 μ ($p=0.0001$).
 μ μ GCS 3-8 μ μ GCS 13-15
 μ .
 μ μ μ μ μ
 μ μ IV (1-1-2-2) μ
 μ μ μ (0.40 (0.20-0.78), $p=0.0058$). Global
 test μ ($p=0.015$, - μ
 Bonferroni 0.008). μ μ IL-1
 (Linkage Disequilibrium) (17) μ
 μ μ D (0,77840) μ μ μ μ
 IL-1 -889C/T IL-1 +3953C/T.
 μ μ IL-2 (IL-2
 +114G/T IL-2 -384T/G) μ μ μ
 μ GCS 3-8 GCS 9-12. , μ μ μ GCS
 3-8 μ μ GCS 13-15 II (2-1)

μ	μ				(0.55 (0.39-0.78),
p=0.00087).	Global test			μ	(p=0.0032).
				(Linkage Disequilibrium)	LD
μ		μ	μ		18

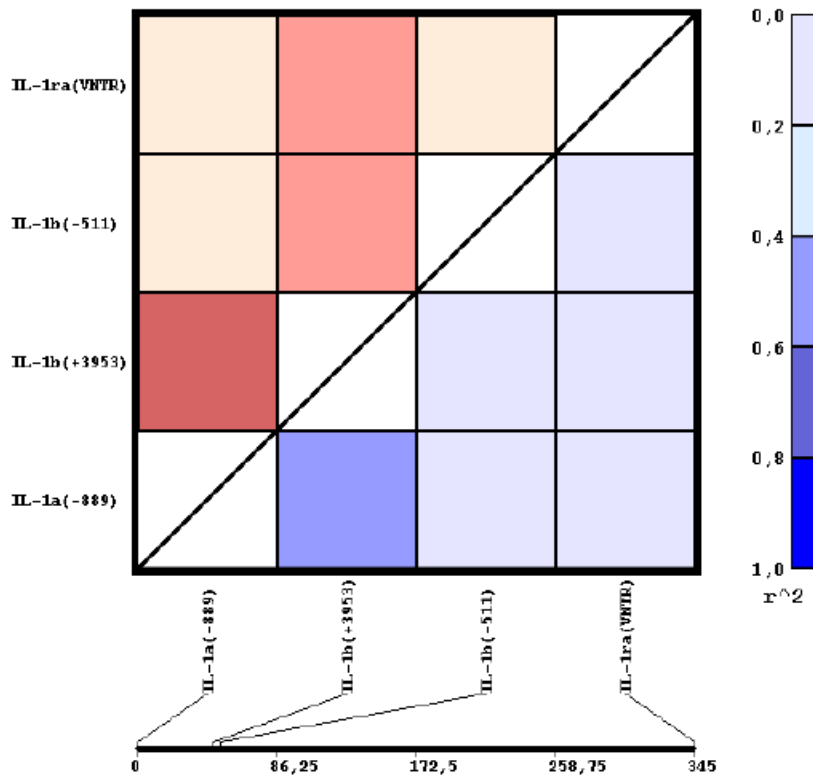
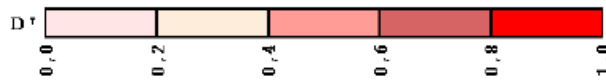
15 (): μ μ IL-1								
Haplotype	IL-1 (-889C/T) rs1800587	IL-1 (+3953C/T) rs1143634	IL-1 (-511C/T) rs16944	IL-1RA (VNTR) rs2234663	% μ GCS 3-8	% μ GCS 13-15	Fisher's p-value	OR (95%CI)*
I	1	1	1	1	25.6	28.1	0.44	1.15 (0.79-1.68)
II	2	2	1	1	15.2	15.4	0.92	1.02 (0.65-1.61)
III	1	1	2	1	14.3	18.2	0.17	1.36 (0.86-2.13)
IV	1	1	2	2	15.0	13.3	0.54	0.86 (0.54-1.38)
V	1	1	1	2	9.1	5.7	0.10	0.59 (0.32-1.11)
VI	2	1	1	1	7.1	5.1	0.31	0.70 (0.36-1.38)
				Global x ² =5.55	Global Fisher's p-value=0.35			
Haplotype	IL-1 (-889C/T) rs1800587	IL-1 (+3953C/T) rs1143634	IL-1 (-511C/T) rs16944	IL-1RA (VNTR) rs2234663	% μ μ	% μ	Fisher's p-value	OR (95%CI)‡
I	1	1	1	1	24.6	34.5	0.08	1.41 (0.95-2.08)
II	2	2	1	1	14.5	18.6	0.46	1.19 (0.74-1.91)
III	1	1	2	1	15.6	19.2	0.59	1.13 (0.71-1.80)
IV	1	1	2	2	13.9	6.9	0.0058	0.40 (0.20-0.78)
V	1	1	1	2	7.9	3.8	0.038	0.40 (0.17-0.97)
VI	2	1	1	1	5.3	8.1	0.31	1.42 (0.71-2.82)
				Global x ² =14.06	Global Fisher's p-value=0.015			

(*) , μ GCS / μ μ μ . (‡) ,

17: (Linkage Disequilibrium, LD) μ -

IL-1RA (VNTR)	0,38520	0,51583	0,34572		
IL-1b (-511)	0,35454	0,42757		0,06464	
IL-1b (+3953)	0,77840		0,03784	0,02978	
IL-1a (-889)		0,43222	0,03655	0,02345	
D'	r^2	IL-1a (-889)	IL-1b (+3953)	IL-1b (-511)	IL-1RA (VNTR)

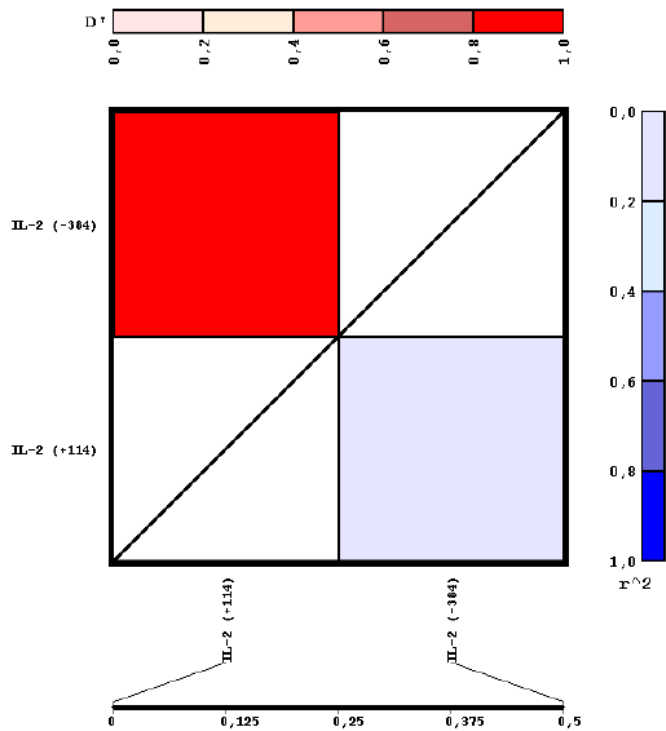
μ : μ μ μ μ $(D' \quad r^2)$ μ
IL-1 (chr 2).



18: *IL-2 (chr 4)* (Linkage Disequilibrium, LD) μ -

<i>IL-2 (-384)</i>	1,00000	
<i>IL-2 (+114)</i>		0,18010
D'	r^2	<i>IL-2 (+114)</i> <i>IL-2 (-384)</i>

μ : μ μ μ *IL-2 (chr 4)*. (D' r^2) μ



μ μ ,
 μ μ μ , μ
 μ
 μ IL-1 , IL-1 , IL-
1RA, IL-2 IL-6, μ μ -
 μ . μ -
 μ μ , μ
 μ μ
 μ .
 μ
IL-1 (-511C/T, rs16944), IL-1 (+3953C/T, rs1143634), IL-1 (-889C/T,
rs1800587), IL-6(-174G/C, rs1800795).
 μ $\mu\mu$ μ μ
IL-1RA VNTR (rs2234663) IL-2 +114G/T (rs2069763)
 μ . μ , VNTR (rs2234663) μ μ IL-1RA
, 6μ
 μ μ . ,
+114G/T (rs2069763), μ μ IL-2
 μ . μ
 μ Bonferroni
 μ 0.007, μ μ .
 μ μ μ .
 μ , IL-1 (IL-1 -889C/T, IL-1 +3953C/T,
IL-1 -511C/T, IL-1RA VNTR) IV (1-1-2-2)
 μ μ μ . ,
VIII (2-2-2-1) VI (2-1-1-1)
 6μ .
 μ μ IL-2 (IL-2 +114G/T IL-2 -384T/G)
II (2-1) μ μ

IL-1RA VNTR (rs2234663)

IL-1: IL-1RA, 1 1 . (VNTR, rs2234663) IL-1RA
 [404].
 IL-1 IL-1
 rs2234663) IL-1RA (VNTR, 2 ()
 4.5 ([4.57 [1.67-12.96], p=0.004).
 (0.26 [0.14-0.71], p=0.003).
 : GCS 3-8 GCS 9-12
 (0.32 [0.12-0.90], p=0.030 GCS 3-8 GCS 13-15
 0.32 [0.16-0.83], p=0.02) GCS 3-8 GCS 13-15
 (0.37 [0.19-0.78], p=0.02 0.45 [0.25-0.91], p=0.017).
 : 0.39 [0.19-0.89], p=0.011
 0.43 [0.19-0.86], p=0.035.

1. IL-1RA VNTR (rs2234663)

VNTR IL-1RA
 IL-1RA
 [322].
 IL-1RA
 IL-1RA
 IL-1RA (sIL-1RA)
 [351, 411].

μ , μ [536].
 μ IL-1RA (icIL-1RA1)
, μ , μ ,
[537]. μ 2 μ μ
 μ , ,
 μ 2 IL-1RA μ μ μ
icIL-1RA1 [404, 412]. ,
 μ IL-1 -
 μ [404].
 μ μ knockout IL-1RA
 μ μ μ μ
[399]. μ , μ
 μ , ,
 μ μ μ -
 μ μ μ μ , μ μ μ -
 μ . μ -
, μ μ μ μ . μ
 μ μ μ μ μ μ μ -
, μ , μ -
 μ IL-1RA
 μ IL-1,
 μ [399]. IL-1RA
 μ μ μ IL-1.
 μ μ μ μ μ μ icIL-1RA1,
 μ μ 2 IL-1RA,
 μ μ μ μ -
 μ [412] μ
 μ 2 2 μ 3 μ icIL-1RA1 μ
 μ μ 1 IL-1RA, icIL-1RA1
 μ . ,
 μ μ μ μ
 μ μ μ IL-1RA

0.91], p=0.017).

GCS 13-15, 2 34,75% IL-1RA 14,58% 20,59% GCS 3-8, GCS 9-12 [71]

(mRNA μ 15min 1h). IL-1

μ IL-1 [542] μ

μ IL-1RA μ IL-1 μ IL-1

μ μ (pmoles) [78]. IL-1 μ μ [85].

μ IL-1RA μ μ μ μ (μ), μ μ 1 [543].

μ μ 4 [544], sIL-1RA μ μ 50% μ μ μ [108].

μ μ IL-1RA μ μ [115],

type), μ (wild
 μ TNF^{-/-}
 μ , μ μ -
 μ IL-1 (μ), TNF
 μ μ ,
 $\mu\mu$ [78]. -
 μ μ IL-1 IL-1 .
 μ IL-1
 . , μ
 ,
 NGF (Nerve Growth Factor) CNTF (Ciliary Neurotrophic Factor).
 μ
 μ . -
 , IL-1 μ .
 μ NGF
 [548]. IL-1 μ ,
 NGF [549, 550]. ,
 CNTF [94] IL-1 . , IL-1RA
 μ μ NGF [551].
 IL-1,
 , IL-1 Ca²⁺ [552]
 μ , GABA [75]
 - [168]. IL-1
 , μ ,
 μ
 [543].
 IL-1 μ μ μ μ
 .
 , μ IL-1 μ μ
 μ
 $\mu\mu$. , μ
 μ ,
 [553, 554] μ

[555]. IL-1 μ μ [95].
 , IL-1 $\mu\mu$
 [556]. μ $\mu\mu$
 Parkinson, μ μ μ
 IL-1 , μ μ
 , μ [557]. IL-1

μ μ μ , μ μ
 μ (long-term potentiation) [558].
 IL-1 μ μ
 μ μ 2 IL-1RA
 μ μ μ .
 μ 2 IL-1RA - μ
 μ μ μ IL-1 μ μ
 icIL-1RA1. , μ 2 IL-1RA μ
 - μ
 μ μ μ μ ,
 μ , μ μ -
 . , μ -
 μ .

μ μ IL-1
 IL-1 (IL-1 -889C/T, IL-1 +3953C/T, IL-1 -511C/T, IL-1RA VNTR), μ -
 μ μ (Linkage Disequilibrium) μ
 μ μ D (0,77840) μ μ μ -
 μ IL-1 -889C/T IL-1 +3953C/T 47
 bp.

μ ,
 IV (1-1-2-2) μ μ
 (0.16 (0.05-0.51), p=0.0005) μ VIII (2-2-2-1)
 μ μ GCS 9-12 (9.79 (2.74-34.98),
 p=2.64*10⁻⁵). μ
 μ 2 VNTR μ μ IL-1RA IV

(1-1-2-2) μ VIII. μ , IV
 μ 2 VNTR μ μ IL-1RA
 μ μ (0.40 (0.20-0.78), p=0.0058). , μ -
 VI (2-1-1-1) μ -
 μ 6 μ μ (
 [μ μ] 3.01 [1.59-5.69], p=0.0004).

μ μ IL-2 +114 (rs2069763) -384
(rs2069762)

IL-2 μ μ IL-2 .
 μ - μ μ
 . μ (natural Killer)
 Th1 (T-helper 1) [346]. -
 IL-2.
 IL-2 μ μ μ
 μ [346]. μ μ μ
 μ IL-2 μ μ
 [444].
 μ μ IL-2 μ μ , μ μ
 -384 [458],
 IL-2 [361] μ
 [461]. O μ μ
 +114 [459]. μ
 μ , μ
 μ μ μ μ
 -384 μ μ .
 μ , μ μ +114G/T (rs2069763) IL-2
 μ 2 () μ μ
 μ , μ
 μ μ μ GCS 3-8 μ GCS 13-15 μ
 μ ([μ μ] 0.57 [0.38-0.80],
 p=0.002) μ μ (0.53 [0.31-0.81], p=0.009).
 μ μ μ μ Bonferroni
 μ μ p=0.02, μ 2.32
 μ μ 1.18 5.12.

μ μ μ GCS 3-8 μ GCS 9-12,
 μ μ μ μ GCS 3-8, GCS 9-12,
 12 GCS 13-15, μ μ μ (3, 143).
 μ 37,20% 31,25% 24,73%
 μ μ μ μ μ -
 μ μ GCS 3-8 μ GCS 9-12.
 1 μ +114G/T (rs2069763) IL-2
 (CTG CTT, Leu Leu 38)
 μ μ mRNA μ -
 μ (Linkage Disequilibrium) μ
 μ μ μ μ ,
 μ μ
 μ μ IL-2 (IL-2 +114G/T IL-2 -384T/G).
 II (2-1) μ μ
 (GCS 3-8 μ GCS 9-12: OR (95%C.I.) 0.55 (0.39-0.78),
 p=0.00087). μ
 μ μ +114G/T μ μ -384T/G.
 μ 1 () μ μ -384T/G μ μ
 IL-2 μ μ 2 (G) [361]. μ μ μ
 μ IL-2 μ μ .
 μ IL-2 μ μ
 (Linkage Disequilibrium) μ
 μ μ LD block (D =1.00 μ
 $r^2=0,18$).

μ μ IL-6 -174G/C (rs1800795)

μ μ μ μ
 μ μ -174G/C (rs1800795) IL-6. IL-6 μ
 μ IL-6 μ , μ -
 μ
 [356].

IL-6 μ μ μ
[559]. , IL-6
, IL-6 [77].
IL-6 μ IL-6 ,
IL-6 μ [560].
 μ μ μ μ μ -174G/C (rs1800795) IL-6.
 μ μ μ IL-6 μ in vivo
[369, 512, 513] μ μ IL-6 [514].
2 : C (), G ()
). CC μ μ IL-6
 μ μ , GG
IL-6 μ [515, 516]. , μ μ
C IL-6 [517],
 μ IL-6 μ CC [518].
 μ μ . μ , μ -
 μ , G μ μ 0.87-
1.0, μ , μ 0.54-0.62 [519-521].
 μ μ -174G/C (rs1800795) IL-6 μ μ -
/
 μ . IL-6,
 μ . , μ μ μ
 μ [272],
 μ μ μ μ .
 μ μ : μ μ $\mu\mu$
, μ μ
 μ μ μ μ
. μ
 μ μ μ .
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, μ .
 μ μ .
 μ μ , μ μ

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 . μ , μ
 μ μ μ , μ
 μ μ μ μ μ
 μ , apoE μ
 μ μ μ μ μ
 apoE μ μ -
 μ bapineuzumab, μ
 μ Alzheimer, μ APOE .
 μ IL-1RA, μ
 μ [535].
 μ ,
 μ ,
 μ μ , μ
 - μ (proinflammatory) - μ (anti-inflammatory) -
 , μ μ μ μ μ .
 μ μ μ μ μ
 μ [561].
 , μ
 μ μ μ μ (genetic
 manipulation), μ μ
 (promoters) [562]. -
 mRNA,
 μ μ [563].
 , μ μ μ μ μ
 μ
 . μ μ (pharmacogenomics)
 μ μ
 μ , μ
 μ μ μ
 [564]. μ μ μ
 μ μ μ μ μ ,
 μ μ μ μ .

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